

Review Article

## Smooth muscle relaxant activity of *Crocus sativus* (saffron) and its constituents: possible mechanisms

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### Abstract

Saffron, *Crocus sativus* L. (*C. sativus*) is rich in carotenoids and used in traditional medicine for treatment of various conditions such as coughs, stomach disorders, amenorrhea, asthma and cardiovascular disorders. These therapeutic effects of the plant are suggested to be due to its relaxant effect on smooth muscles. The effect of *C. sativus* and its constituents on different smooth muscles and the underlying mechanisms have been studied. Several studies have shown the relaxant effects of *C. sativus* and its constituents including safranal, crocin, crocetin and kaempferol on blood vessels. In addition, it was reported that saffron stigma lowers systolic blood pressure. The present review highlights the relaxant effects of *C. sativus* and its constituents on various smooth muscles. The possible mechanisms of this relaxing effect including activation of  $\beta_2$ -adrenoceptors, inhibition of histamine  $H_1$  and muscarinic receptors and calcium channels and modulation of nitric oxide (NO) are also reviewed.

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### Introduction

*Crocus sativus* L (*C. sativus*), commonly known as saffron, is a small perennial plant belonging to the family of Iridaceae which is cultivated in many countries with mild and dry climate specially in Iran (Abdullaev, 1993). It has gray-green hairy margins leaves and grows to about 30-35 cm long and has a funnel-shaped reddish purple flower with red stigmas. The stigmas are commonly used as a flavoring and yellow coloring additive in various

foods such as bread, soups, sauces and rice (Kataria et al., 2011).

Main constituents of saffron stigmas are crocin (responsible for its color), picrocrocin (responsible for its bitter taste) and safranal (responsible for its odor and aroma). It also contains more than 150 volatile aroma-yielding compounds and many non-volatile active components, many of which are carotenoids including zeaxanthin, lycopene and various alpha- and beta-carotenes (Srivastava et al., 2010).

In folk medicine, *C. sativus* has been used as an antispasmodic, eupeptic, gingival sedative, anticatarrhal, sedative, carminative, diaphoretic, expectorant, stimulant, stomachic, aphrodisiac and emmenagogue medicine (Nemati *et al.*, 2008).

There also appears to be adequate evidence to support therapeutic effects of saffron and also for its use in alleviating several health disorders.

Recent studies showed pharmacological effects of saffron such as antioxidant (Hosseinzadeh *et al.*, 2009; Soeda *et al.*, 2007) antitumor (Abdullaev, 2002; Aung *et al.*, 2007; Mousavi *et al.*, 2011), memory enhancing (Abe and Saito, 2000; Ghadrdoost *et al.*, 2011; Hosseinzadeh and Ziaei, 2006; Pitsikas *et al.*, 2007), antidepressant and anxiolytic (Hosseinzadeh *et al.*, 2003; Hosseinzadeh and Noraei, 2009), genoprotective (Hosseinzadeh and Sadeghnia, 2007; Premkumar *et al.*, 2001), antitussive (Hosseinzadeh and Ghenaati, 2006), cardioprotective (Goyal *et al.*, 2010; Imenshahidi *et al.*, 2010; Xu *et al.*, 2006; Zhang *et al.*, 2009) and neuroprotective activities (Essa *et al.*, 2012; Mehri *et al.*, 2012).

Some of saffron therapeutic effects on stomach disorders, colic, dysmenorrhoea, asthma and cardiovascular disorders are suggested to be due to its relaxant effect on smooth muscles. A potent relaxant effect of the plant extract on tracheal smooth muscle was shown which was comparable to theophylline and greater than the effect of its constituent, safranal (Boskabady and Aslani, 2006).

The effects of saffron petals extracts on blood pressure, hypotensive effect of aqueous extract of *C. sativus* and its constituents, safranal and crocin (Fatehi *et al.*, 2003), effects of chronic and subchronic crocin treatment on hypertension (Imenshahidi *et al.*, 2014; Razavi *et al.*, 2013) and effect of chronic administration of saffron stigma aqueous extract on systolic blood pressure were shown (Imenshahidi *et al.*, 2013). The

protective effect of crocin on reperfusion-induced cardiac arrhythmias (Jahanbakhsh *et al.*, 2012) as well as the lowering effect on heart rate and contractility was also documented (Boskabady *et al.*, 2008). In addition, the effect of saffron on uterine contraction was also investigated (Chang *et al.*, 1964). For *C. sativus* and its constituent safranal, a stimulatory effect on  $\beta$ -adrenoceptors (Nemati *et al.*, 2008), an inhibitory effect on histamine H<sub>1</sub> receptors (Boskabady *et al.*, 2010) and a functional antagonistic effect on muscarinic receptors were demonstrated (Neamati and Boskabady, 2010).

To prepare the present review, we explored the literature regarding the smooth muscle relaxant effect of *C. sativus* and its constituent as well as the possible underlying mechanisms.

## Methods

The related data on this topic was obtained by searching for the terms "*Crocus sativus*", "saffron", "crocin", "crocin", "safranal", "smooth muscles" and "relaxant effects" using ISI Web of Knowledge, Medline/Pubmed, Science direct, Scopus, Google Scholar, Embase, Biological Abstracts and Chemical Abstracts.

## Constituents

More than 150 compounds have been extracted from saffron stigmas, from which less than 50 chemicals have been completely characterized. Among fully characterized compounds, some are colored carotenoids e.g. crocetin and its glycosidic derivatives, crocins. The other components are colorless monoterpene aldehydes, volatile agents e.g. safranal, and bitter components, e.g. picrocrocin. The most important components are listed in a review article by Bathaie and Mousavi (Bathaie and Mousavi, 2010).

Safranal (fat soluble) and pigments of the crocetin carotenoid (a natural dicarboxylic acid carotenoid precursor

crocin) are bitter, but the most important cause of saffron bitterness is picrocrocin (Abdullaev, 1993). Color compounds of saffron are crocetin carotenoids and glycosidic forms of di-gentiobioside (crocin), gentiobioside, glycoside, gentio-glycoside and beta-crocetin di-glycoside (mono-methyl ester), gama-crocetin (di-methyl ester), alpha-carotene, beta-carotene, lycopene and zeaxanthin. Saffron lipophilic carotenoids are lycopene, alpha- and beta-carotene and zeaxanthin (Winterhalter and Straubinger, 2000; Tarantilis and Polissiou, 1997). Kampferol has also been found in alcoholic extract of saffron petals (Gregory et al., 2005). Flavonoids, especially lycopene, amino acids, proteins, starch, resins and other compounds have also been found in saffron (Assimopoulou et al., 2005). Moreover, saffron has trace amounts of thiamine and riboflavin (Alonso et al., 2001).

The main constituents of saffron are crocetin and its digentiobiosyl ester, crocin. Traces of non-glycosylated carotenoids unrelated to crocetin like  $\beta$ -carotene, lycopene and zea-xanthin are also present (Rios et al., 1996). Ethanolic extract of saffron has visible absorption peaks at 427 and 452 nm. When excited at 435 nm, saffron emits at 543 nm (Horobin et al., 2002). The water soluble extract of saffron stigmas is intensely colored; even one part of saffron in 100,000 parts of water yields a visibly yellow solution. The color components of saffron are diapocarotenoids, which are chemically similar to those in annatto.

Crocetin, another carotenoid that is isolated from saffron is one of the two principal chemicals responsible for saffron red color (Martin et al., 2002). Crocetin, a diapocarotenoid, is the result of crocin glycosides hydrolysis. Crocetin contains a carboxyl group at each end of the polyene chain; when ionized, it can function as an acid (anionic) dye for biological staining (Lillie, 1977). Crocetin in its free-acid form is insoluble in water and most organic solvents, except for pyridine and

dimethylsulfoxide. Its anionic species are highly water soluble, so crocetin readily dissolves in dilute aqueous sodium hydroxide or other aqueous alkali solutions  $\text{pH} \geq 9$ . Absorption maxima in pyridine are at 411, 436 and 464 nm. Crocetin constitutes approximately 0.3% of the total weight of the saffron stigma (Escribano et al., 1996; Dris and Jain, 2004). Crocetin is present mostly as its trans isomer. On the other hand, cis-crocetin and its glycosides are only minor components of saffron.

Crocetin (digentiobiosyl 8,8'-diapocarotene-8,8'-oate) belongs to a group of natural carotenoid commercially obtained from the dried stigma of saffron. It is the diester that is formed from the disaccharide gentiobiose and the dicarboxylic acid crocetin.

Crocins account for almost 6–16% of saffron dry weight (based on its variety, cultivation environment and type of processing) (Gregory et al., 2005). Crocin (crocin 1) is a digentiobioside carotenoid which comprises the majority of crocins found in saffron. Due to its sugar parts, crocin could be easily dissolved in water, turning from a dark red dry powder to an orange aqueous solution making saffron a desirable coloring food additive (Melnyk et al., 2010). Its deeply red colored crystals have a melting point of 186°C.

Crocetin structure was elucidated by Karree (Karrer et al., 1933) though its presence was reported by Aschoff in the 19th century, as saffron main pigment (approx. 80%). Using water as the stationary phase and butanol as the mobile phase, crocetin can be isolated from saffron extract in pure form and is directly crystallized.

Oral administration of single or multiple doses of crocetin is not absorbed through the gastrointestinal tract and is largely excreted. It has been observed that orally administered crocins are hydrolyzed to crocetin before or during intestinal absorption, and absorbed crocetin is partly metabolized to mono- and diglucuronide conjugates (Asai et al., 2005). Crocetin does not tend to accumulate following

administration of repeated oral doses and the intestinal tract serves as an important site for crocin hydrolysis (Xi *et al.*, 2007).

### **Relaxant effect of *C. sativus* and its constituents on different types of smooth muscles**

The relaxant effects of *C. sativus* extracts and constituents on various smooth muscles are reviewed in the following sections.

#### **Vascular smooth muscle**

Imenshahidi *et al.* compared the hypotensive effect of saffron aqueous extract and its two active ingredients in rats. Based on the results, aqueous extract of saffron stigma, safranal and crocin decreased mean arterial blood pressure in a dose-dependent manner. The hypotensive effect of the extract is perhaps due to its relaxant effect on vascular smooth muscle. The results also suggested that safranal, the major constituent of the plant, contributes to the hypotensive activity (Imenshahidi *et al.*, 2010).

Fatehi *et al.* investigated the effects of *C. sativus* petals extract on blood pressure in anesthetized rats. Aqueous and ethanolic extracts of *C. sativus* petals reduced blood pressure in a dose-dependent manner. Administration of 50 mg/g of aqueous extract reduced blood pressure from  $133.5 \pm 3.9$  to  $117 \pm 2.1$  mmHg. This hypotensive effect could be either due to the effect of the *C. sativus* petals extracts on the heart itself or on total peripheral resistance via relaxation of vascular smooth muscle, or both. However, the results suggested that the effect of extracts on peripheral resistance seems to be more probable mechanism of this effect (Fatehi *et al.*, 2003).

It was also shown that chronic administration of aqueous extract of saffron stigma reduced DOCA-induced increase in mean systolic blood pressure (MSBP), but this hypotensive effect was not observed in normotensive rats. Data showed that

antihypertensive effects of saffron did not last for a long period, so it could be postulated that long term blood pressure regulation systems were not affected by saffron (Imenshahidi *et al.*, 2013).

Aqueous-ethanolic extract of *C. sativus* also showed concentration-dependent inhibitory effect on heart rate and contractility comparable to the effect of diltiazem. The effect of plant extract on heart contractility could be due to its muscle relaxant effect (Boskabady *et al.*, 2008).

It was also shown that crocetin (15, 30 mg/kg) dose-dependently improved endothelium-dependent relaxation (EDR) in response to acetylcholine (ACh) in high cholesterol diet (HCD)-fed rabbits. Also, in bovine aortic endothelial cells (BAECs), oxidized LDL (oxLDL) treatment decreased nitric oxide production and downregulated the activity and mRNA expression of endothelial nitric oxide synthase but these effects were inhibited by crocetin (0.1, 1, 10 mM) in a dose-dependent manner (Tang *et al.*, 2006).

The effect of crocin (50 mg/kg) on improvement of the reduction of systolic blood pressure (SBP) and the increased heart rate (HR) induced by diazinon (DZN) in rats, was shown which could be due to the relaxant effect of crocin on muscle cells (Razavi *et al.* in press).

The vasodilator effects of quercetin and its metabolites on vessels resistance were also documented (Xu *et al.*, 2015). The relaxant effect of kaempferol, the other constituent of *C. sativus* was also examined in several studies. In one study, low concentration of kaempferol (10  $\mu$ M) caused endothelium-dependent and endothelium-independent vascular relaxations (Xu *et al.*, 2006). In addition, relaxant effect of kaempferol on smooth muscle cells of the porcine coronary artery was demonstrated (Xu *et al.*, 2006). It has been also reported that quercetin and kaempferol relaxed rat aortic strips contraction induced by noradrenaline, KCl or phorbol 12-myristate,13-acetate (Duarte *et al.*, 1993).

### Tracheal smooth muscle

In a study by Boskabady and Aslani (2006), on guinea-pig tracheal smooth muscle, it was shown that aqueous-ethanolic extract of saffron has a potent relaxant effect, which was comparable to theophylline. Also, it was observed that there were positive correlations between increasing concentrations and the relaxant effects of the extract (Boskabady and Aslani, 2006).

Safranal also showed concentration-dependent relaxant effect on tracheal smooth muscle. However, the relaxant effect of safranal was lower than that of the extract and theophylline (Boskabady and Aslani, 2006). These results indicated that relaxant effect of *C. sativus* is in part due to its constituent safranal.

Long term oral administration of *C. sativus* extract (Bayrami et al., 2013) and its constituent safranal (Boskabady and Byrami, 2014) also reduced tracheal responsiveness to methacholine in sensitized guinea pigs. This effect could be due to relaxant effect of the extract and safranal on tracheal smooth muscle. The antitussive effect of the ethanolic extract of saffron and safranal was shown which could be due to their relaxant effect on airway smooth muscle (Hosseinzadeh and Ghenaati, 2006).

### Gastrointestinal and urogenital smooth muscle

The effects of *C. sativus* petals extracts on isolated guinea-pig ileum induced by electrical field stimulation (EFS) was studied. In rat isolated ileum, contractile responses to EFS were decreased by the petals extracts. Contractions of ileum to EFS are mediated by both noradrenaline and ATP released as co-transmitters from sympathetic nerves (Hoyle and Burnstock, 1991). The results showed that ethanolic extract had a greater relaxant effect on EFS-induced contraction of guinea pig ileum than the aqueous extract (Fatehi et al., 2003).

The relaxant effect of saffron on uterine contraction was also demonstrated (Chang et al., 1964). The relaxant effect of *C.*

*sativus* and its constituents on different types of smooth muscles was summarized in Table 1.

### Possible mechanisms underlying smooth muscle relaxant effect of *C. sativus*

#### *β<sub>2</sub> – adrenoreceptors stimulatory effect*

The most possible mechanism for the relaxant effect of agents on tracheal smooth muscle is their stimulatory effect on  $\beta_2$ -adrenergic receptors. The  $\beta_2$ -adrenergic stimulatory effect of the plant and safranal was tested by performing cumulative concentration-response curves of isoprenaline-induced relaxation of pre-contracted isolated guinea pig tracheal smooth muscle. In this regard, aqueous ethanolic extracts from *C. sativus* (0.1 and 0.2 %), safranal (1.25 and 2.5  $\mu$ g), 10 nM propranolol and saline were studied. The results showed clear leftward shifts in isoprenaline curves obtained in the presence of saffron extract and safranal compared to that of saline while propranolol caused rightward shift in isoprenaline response curve. The results indicated a relatively potent stimulatory effect for *C. sativus* extract and its constituent safranal on  $\beta_2$ -adrenoreceptors (Nemati et al., 2008). Therefore, the results suggested that the major mechanism responsible for the relaxant effect of the plant and safranal is their stimulatory effect on  $\beta_2$ -adrenoreceptors.

#### *Anti-cholinergic and anti-muscarinic effect of the plant*

The study of Nemati et al. (2010) demonstrated the functional antagonistic effect of *C. sativus* and safranal on muscarinic receptors on tracheal muscle of guinea pigs. Both the extract and safranal shifted methacholine concentration-response curve to the right. However, the shift was not parallel and the maximum response to methacholine in the presence of extract and safranal was not obtained. These results indicated a functional antagonistic effect of the plant and safranal on muscarinic receptors (Nemati and Boskabady, 2010).

Table 1. The relaxant effect of *C. sativus* and its constituents on different types of smooth muscle

Type of smooth muscle (SM)	Solution	Effect	Reference
Vascular	AE of saffron stigma, safranal and crocin	Decreased the mean arterial blood pressure of animals	Imenshahidi et al., 2010
	AE and EE of <i>C. Sativus</i> petals	Reduced the blood pressure in a dose-dependent manner	Fatehi et al., 2003
	AE of <i>C. Sativus stigma</i>	Reduced the enhanced of mean systolic blood pressure	Imenshahidi et al., 2013
	AE and EE of <i>C. Sativus</i>	Inhibitory effect on heart rate and contractility	Boskabady et al., 2008
	Crocetin, (15, 30 mg/kg)	Improved endothelium-dependent relaxation (EDR) in response to acetylcholine	Tang et al., 2006
	Crocetin (50 mg/kg)	Crocetin (50 mg/kg) and DZN in improvement of the reduction of SBP and the elevation of HR	Razavi et al. In press
Trachea	AE and EE of <i>C. Sativus</i>	Potent relaxing effect on the tracheal smooth muscle.	Boskabady et al., 2006
	Safranal	Has relaxant effect on tracheal smooth muscle	Boskabady and Aslani, 2006
	<i>C. Sativus</i>	Reduction effect on tracheal responsiveness to methacholine in animals	Bayrami et al., 2013
	Safranal	Reduction effect on tracheal responsiveness to methacholine in animals	Boskabady and Byrami, 2014
Gastrointestinal	AE of saffron and safranal	Antitussive effect was shown	Hosseinzadeh and Ghenaati, 2006
	Extract of <i>C. Sativus</i> petals	Induced relaxant effect in guinea -pig ileum	Fatehi et al., 2003
	Saffron	Relaxant effect on uterine contraction	Chang et al., 1964

Aqueous extracts (AE), ethanolic extracts (EE)

#### *Histaminic (H<sub>1</sub> receptor) antagonistic activity*

The effect of three concentrations of aqueous-ethanolic extracts of *C. sativus* (0.025 %, 0.05 % and 0.1 %) on histamine (H<sub>1</sub>) receptors was evaluated in guinea pig tracheal smooth muscles. Concentration-response curve for histamine was obtained in the presence of saline, saffron extract and chlorpheniramine. The extract caused parallel rightward shift in histamine concentration-response curve similar to the effect of chlorpheniramine and the maximum response to histamine was obtained in the presence of the extract. These results showed an inhibitory effect of *C. sativus* (competitive antagonistic effect) on histamine H<sub>1</sub> receptors which could be related to the relaxant effect of the plant on

tracheal smooth muscle (Boskabady et al., 2010).

The effect of safranal (0.63, 1.25 and 2.5 µg/ml) on histamine (H<sub>1</sub>) receptors in guinea pig tracheal smooth muscle was also studied using similar method as described above. The results showed that safranal also caused rightward shift in histamine-response curves, obtaining the maximum responses to histamine and greater EC<sub>50</sub> (effective concentration of histamine causing 50% of maximum response). It was concluded that safranal possibly acts as a histamine H<sub>1</sub> receptors competitive antagonist (Boskabady et al., 2011).

#### *Calcium channel antagonistic effects*

Crocetin could inhibit Ca<sup>2+</sup> influx and release of intracellular Ca<sup>2+</sup> stores in the endoplasmic reticulum in bovine aortic

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smooth muscle cells (He et al., 2004). It was also shown that reduction of intracellular  $\text{Ca}^{2+}$  release may contribute to relaxation of the corpus cavernosum, leading to erection (Williams et al., 2005). Coronary and other diseases in cardiac or brain blood vessels are considered to be due to the excessive influx of  $\text{Ca}^{2+}$  into cytoplasm. So,  $\text{Ca}^{2+}$  channels blockers are of therapeutic value in treatment of these diseases. The effect of *C. sativus* on  $\text{Ca}^{2+}$  influx in isolated rat aortas was investigated using  $^{45}\text{Ca}$  as a radioactive tracer, and their calcium antagonistic effects were evaluated.  $\text{Ca}^{2+}$  uptake in isolated rat aorta rings in normal physiological status was not markedly altered by these drugs, whereas  $\text{Ca}^{2+}$  influx induced by norepinephrine 1.2  $\mu\text{mol/L}$  and KCl 100  $\text{mmol/L}$  were significantly inhibited by *C. sativus* in a concentration-dependent manner.

The results showed that  $\text{Ca}^{2+}$  influx through receptor-operated  $\text{Ca}^{2+}$  channels and potential-dependent  $\text{Ca}^{2+}$  channels can be blocked by the plant (Liu et al., 2005). It is conceivable that the hypotensive effect of saffron in chronic treatment is related to its inhibitory effect on smooth muscles via blocking calcium channel or inhibiting sarcoplasmic reticulum  $\text{Ca}^{2+}$  release into the cytosol (Boskabady and Aslani, 2006).

Crocic acid also concentration-dependently inhibited the total cholesterol (TC) and Cholesteryl ester (CE) elevation induced by Ox-LDL. The results indicated that crocin could inhibit the increased intracellular [ $\text{Ca}^{2+}$ ] in smooth muscle cell (He et al., 2005).

Concerning the underlying mechanisms of the vasodilatory effect, protein kinase C inhibition and decrease in  $\text{Ca}^{2+}$  uptake were also investigated (Duarte et al., 1993).

It has been reported that crocetin decreased protein kinase C (PKC) activity in the membrane fraction, which led to reduced blood pressure by inhibition of proliferation in vascular smooth muscle cells (Cheng-Hua et al., 2010).

### *Endothelium-dependent relaxation (EDR) effect*

The mechanisms by which crocetin causes vasorelaxation might be attributed to the vessel endothelial instead of direct effect on vessel smooth muscle. It was shown that crocetin upregulated the eNOS mRNA expressions in both in vivo and in vitro studies which was in accordance with its action on eNOS activity and NO production. The NO formed in the endothelial cell is catalyzed by eNOS, crosses the plasma membrane and diffuses into the adjacent smooth muscle cells, where it binds and stimulates guanylyl cyclase, the enzyme that synthesizes cGMP. Cyclic GMP leads to a decrease in cytosolic  $\text{Ca}^{2+}$  concentration, which causes relaxation of the muscle cell and dilation of the blood vessel (Tang et al., 2006).

Smooth muscle cells relaxations of the porcine coronary artery by endothelium-derived and exogenous NO due to endothelium-dependent hyperpolarization was shown. These results showed the involvement of activation of large-conductance calcium-activated potassium channel  $\text{K}_{\text{Ca}1.1}$  channels and NO in vascular smooth muscle relaxant effect (Xu et al., 2005). However, further studies should investigate this mechanism to justify the relaxant effect of *C. sativus* and its constituents.

### *Increasing intracellular cAMP*

It was shown that rat uterine smooth muscle relaxation following KCl-induced tonic contraction was antagonized by Rp-cAMPS which indicates involvement of increased intracellular cAMP concentration in relaxing uterine smooth muscle (Revuelta et al., 2000). This mechanism should also be examined for *C. sativus* and its constituents in further experiments.

The possible mechanisms of the relaxant effect of *C. sativus* and its constituents were summarized in Table 2.

Table 2. Possible mechanisms of the relaxant effect of *C. sativus* and its constituents

Possible mechanisms	Solution	Type of muscle	Reference
Stimulatory effect on $\beta_2$ -adrenoceptors	AE extract of <i>C. Sativus</i> (0.1 and 0.2 g%) and safranal	Trachea	(Nemati et al., 2008)
Anticholinergic and anti-muscarinic effect	AE extract of <i>C. Sativus</i> and safranal	Trachea	(Neamati and Boskabady, 2010)
Histaminic antagonistic activity	AE extract of <i>C. Sativus</i> (0.025 g%, 0.05 g% and 0.1 g%)	Trachea	(Boskabady et al., 2010)
	Safranal (0.63, 1.25 and 2.5 $\mu$ g/ml)	Trachea	Boskabady et al., 2011
Ca <sup>2+</sup> channel blocking in aortic smooth muscle cells	Crocine	Vascular	He et al., 2004
Ca <sup>2+</sup> channel blocking in corpus cavernosum leading to erection	Crocine	Vascular	Williams et al., 2005
Protein kinase C inhibition or decreased Ca <sup>2+</sup> uptake	Kaempferol	Vascular	Duarte et al., 1993
Protein kinase C inhibition in the membrane fraction	Crocetin	Vascular	Cheng-Hua et al., 2010
Upregulation of eNOS mRNA expressions and NO production	Crocetin	Vascular	(Tang et al., 2006)
Activation of large-conductance calcium-activated potassium channel $k_{CaV1.1}$ channels and NO	Kaempferol	Vascular	Xu et al., 2015
Increase in intracellular cAMP	Kaempferol	Vascular	Revuelta et al., 2000

## Conclusion

In conclusion, the relaxant effects of *C. sativus* and some of its constituents on vascular, trachea, gastrointestinal and urogenital smooth muscles were discussed in the present review article. The possible mechanisms of the relaxant effect of the plant and its constituents on smooth muscle including  $\beta_2$ -adrenoreceptors stimulation, histamine (H<sub>1</sub>) receptor inhibition and calcium channel blocking were also reviewed. However, the point that exactly which chemical(s) through what mechanism(s) of action causes this effect as well as its clinical applications should be investigated in further studies.

## References:

- Abdullaev FI. 1993. Biological effects of saffron. *BioFactors*, 4: 83–86.
- Abdullaev FI. 2002. Cancer chemopreventive and tumoricidal properties of saffron (*Crocus sativus L.*). *Exp Biol Med*, 227: 20–25.
- Abe K, Saito H. 2000. Effects of saffron extract and its constituent crocin on learning behaviour and long-term potentiation. *Phytother Res*, 14: 149–152.
- Alonso GL, Salinas M R, Garijo J, Sánchez-Fernández M A. 2001. Composition of crocins and picrocrocin from Spanish saffron (*Crocus sativus L.*). *J food qual*, 24: 219–233.
- Asai A, Nakano T, Takahashi M, Nagao A. 2005. Orally administered crocetin and crocins are absorbed into blood plasma as crocetin and its glucuronide conjugates in mice. *J Agric Food Chem*, 53: 7302–6.
- Assimopoulou AN, Sinakos Z, Papageorgiou VP. 2005. Radical scavenging activity of *Crocus sativus L.* extract and its bioactive constituents. *Phytother Res*, 19: 997–1000.
- Aung HH, Wang CZ, Ni M, Fishbein A, Mehendale SR, Xie JT, Shoyama CY, Yuan CS. 2007. Crocin from *Crocus sativus* possesses significant anti-proliferation effects on human colorectal cancer cells. *Exp oncol*, 29: 175–180.
- Bathaie SZ, Mousavi SZ. 2010. New applications and mechanisms of action of saffron and its important ingredients. *Crit Rev Food Sci*, 50: 761–786.
- Bayrami G, Boskabady MH, Jalali S, Farkhondeh T. 2013. The effect of the extract of *Crocus sativus* on tracheal responsiveness and plasma levels of IL-4, IFN- $\gamma$ , total NO and nitrite in ovalbumin sensitized Guinea-pigs. *J EthnoPharmacol*, 147: 530–535.



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- Boskabady M H, Shafei MN, Shakiba A, Sefidi HS. 2008. Effect of aqueous-ethanol extract from *Crocus sativus* (saffron) on guinea-pig isolated heart. *Phytother Res*, 22: 330–334.
- Boskabady MH, Ghasemzadeh M, Nemati H, Esmailzadeh M. 2010. Inhibitory effect of *Crocus sativus* (saffron) on histamine (H<sub>1</sub>) receptors of guinea pig tracheal chains. *Pharmazie*, 65: 300–305.
- Boskabady MH, Aslani MR. 2006. Relaxant effect of *Crocus sativus* (saffron) on guinea-pig tracheal chains and its possible mechanisms. *J Pharm Pharmacol*, 58: 1385–1390.
- Boskabady MH, Byrami G, Faizpour A. 2014. The Effect of Safranal, a Constituent of *Crocus sativus* (Saffron), on Tracheal Responsiveness, Serum Levels of Cytokines, Total NO and Nitrite in Sensitized Guinea Pigs. *Pharmacol Rep*, 66: 56–61.
- Boskabady MH, Rahbardar MG, Jafari Z. 2011. The effect of safranal on histamine (H<sub>1</sub>) receptors of guinea pig tracheal chains. *Fitoterapia*, 82: 162–167.
- Chang PY, Wang CK, Liang JD, Kuo W. 1964. Studies on the pharmacological action of Zang Hong Hua (*Crocus sativus* L.). I. Effects on uterus and estrus cycle. *Yao Xue Xue Bao*, 11: 94.
- Cheng-Hua Zhou, Min Xiang, Shu-Ying He, Zhi-Yu Qian. Protein Kinase C Pathway is Involved in the Inhibition by Crocetin of Vascular Smooth Muscle Cells Proliferation. *Phytother. Res.* 24: 1680–1686 (2010)
- Dris R, Jain SM. 2004. Production Practices and Quality Assessment of Food Crops: Preharvest Practice. Kluwer, Berlin. Volume 1; 218.
- Duarte Juan, Francisco Pi~rez Vizcafno, Pilar Utrilla, Josi~ Jimi~Nez, Juan Tamargo, Antonio Zarzuelo. Vasodilatory effects of flavonoids in rat aortic smooth muscle structure-activity relationships. *Gen. Pharmac.* Vol. 24, No. 4, pp. 857-862, 1993
- Escribano J, Alonso GL, Coca-Prados M, Fernandez JA. 1996. Crocin, safranal and picrocrocin from saffron (*Crocus sativus* L.) inhibit the growth of human cancer cells in vitro. *Cancer lett*, 100:23–30.
- Essa MM, Vijayan RK, Castellano-Gonzalez G, Memon MA, Braidy N, Guillemin GJ. 2012. Neuroprotective effect of natural products against Alzheimer's disease. *Neurochem res*, 37:1829–1842.
- Fatehi M, Rashidabady T, Fatehi-Hassanabad Z. 2003. Effects of *Crocus sativus* petals' extract on rat blood pressure and on responses induced by electrical field stimulation in the rat isolated vas deferens and guinea-pig ileum. *J ethnopharmacol*, 84:199–203.
- Ghadroost B, Vafaei AA, Rashidy-Pour A, Hajisoltani R, Bandegi AR, Motamedi F, Haghighi S, Sameni HR, Pahlvan S. 2011. Protective effects of saffron extract and its active constituent crocin against oxidative stress and spatial learning and memory deficits induced by chronic stress in rats. *Eur J Pharmacol*, 667: 222–229.
- Goyal SN, Arora S, Sharma AK, Joshi S, Ray R, Bhatia J, Kumari S, Arya DS. 2010. Preventive effect of crocin of *Crocus sativus* on hemodynamic, biochemical, histopathological and ultrastructural alterations in isoproterenol-induced cardiotoxicity in rats. *Phytomedicine*, 17: 227–232.
- Gregory MJ, Menary RC, Davies NW. 2005. Effect of drying temperature and air flow on the production and retention of secondary metabolites in saffron. *J Agric Food Chem*, 53:5969–5975.
- He S, Qian Z, Tang F. 2004. Effect of crocin on intracellular calcium concentration in cultured bovine aortic smooth muscle cells. *Acta Pharm Sin*, 39: 778–781.
- He SY, Qian ZY, Tang FT, Wen N, Xu GL, Sheng L. Effect of crocin on experimental atherosclerosis in quails and its mechanisms. *Life Sci.* 2005; 77(8): 907-21.
- Horobin RW, Kiernan JA, Eds. 2002. *Conn's Biological Stains: a Handbook of Dyes, Stains and Fluorochromes for Use Biology and Medicine*, 10<sup>th</sup> ed. BIOS Scientific Publishers, Oxford, UK; 315.
- Hosseinzadeh H, Ziaei T. 2006. Effects of *Crocus sativus* stigma extract and its constituents, crocin and safranal, on intact memory and scopolamine-induced learning deficits in rats performing the Morris water maze task. *J Med Plants*, 5:40–50.
- Hosseinzadeh H, Shamsaie F, Mehri, S. 2009. Antioxidant activity of aqueous and ethanolic extracts of *Crocus sativus* L. stigma and its bioactive constituents, crocin and safranal. *Pharmacog Mag*, 5: 419.

- Hosseinzadeh H, Ghenaati J. 2006. Evaluation of the antitussive effect of stigma and petals of saffron (*Crocus sativus*) and its components, safranal and crocin in guinea pigs. *Fitoterapia*, 77: 446–448.
- Hosseinzadeh H, Karimi G, Niapoor M. 2004. Antidepressant effect of *Crocus sativus* L. stigma extracts and their constituents, crocin and safranal, in mice. *J Med Plants*, 3: 48–58.
- Hosseinzadeh H, Noraei NB. 2009. Anxiolytic and hypnotic effect of *Crocus sativus* aqueous extract and its constituents, crocin and safranal, in mice. *Phytother Res*, 23:768–774.
- Hosseinzadeh H, Sadeghnia HR. 2007. Effect of safranal, a constituent of *Crocus sativus* (saffron), on methyl methanesulfonate (MMS)-induced DNA damage in mouse organs: an alkaline single-cell gel electrophoresis (comet) assay. *DNA and cell biol*, 26:841–846.
- Hoyle CHV, Burnstock G. 1991. ATP receptors and their physiological roles. In: Stone, TW. (Ed.), *Adenosine in the Nervous System*. Academic Press, London, 43-76.
- Imenshahidi M, Razavi B M, Faal A, Gholampoor A, Mousavi S M, Hosseinzadeh H. 2014. Effects of chronic crocin treatment on desoxycorticosterone acetate (doca)-salt hypertensive rats. *Iran J Basic Med Sci*, 17:9.
- Imenshahidi M, Razavi B M, Faal A, Gholampoor A, Mousavi S M, Hosseinzadeh H. 2013. The Effect of Chronic Administration of Saffron (*Crocus sativus*) Stigma Aqueous Extract on Systolic Blood Pressure in Rats. *Jundishapur J Nat Pharm Prod*, 8:175.
- Imenshahidi M, Hosseinzadeh H, Javadpour Y. 2010. Hypotensive effect of aqueous saffron extract (*Crocus sativus* L.) and its constituents, safranal and crocin, in normotensive and hypertensive rats. *Phytother Res*, 24: 990–994.
- Jahanbakhsh Z, Rasouljan B, Jafari M, Shekarforoush Sh, Esmailidehaj M, Mohammadi M T, Aghai H, Salehi M. 2012. Protective effect of crocin against reperfusion-induced cardiac arrhythmias in anaesthetized rats. *EXCLI J*, 11:20–29.
- Karrer P, Benz F, Stoll M. 1933. Pflanzenfarbstoffe XLIX. Synthese des Perhydro-crocetins. *Helv Chim Acta*, 16: 297–302.
- Kataria D, Kumar C, Nerkar N, Gadiya RV, Abhyankar MM. 2011. Detail profile of *Crocus sativus*. *International Journal of Pharma and Bio Sciences*; 2: 530-540.
- Lillie RD. 1977. *Conn's Biological Stains: a Handbook on the Nature and Uses of the Dyes Employed in the Biological Laboratory*, 9<sup>th</sup> ed. Williams & Wilkins, Baltimore, MD; 459.
- Liu N, Yang Y, Mo S, Liao J, Jin J. 2005. Calcium antagonistic effects of Chinese crude drugs: Preliminary investigation and evaluation by <sup>45</sup>Ca. *Appl Radiat Isot*, 63: 151–155.
- Martin G, Goh E, Neff AW. 2002. Evaluation of the developmental toxicity of crocetin on *Xenopus*. *Food Chem Toxicol*, 40: 959–964.
- Mehri S, Abnous Kh, Mousavi SH, Shariaty VM, Hosseinzadeh H. 2012. Neuroprotective effect of crocin on acrylamide-induced cytotoxicity in PC12 cells. *Cell mol neurobiol*, 32: 227–235.
- Melnyk JP, Wang S, Marcone MF. 2010. Chemical and biological properties of the world's most expensive spice: Saffron. *Food Chem Toxicol*, 43: 1981–1989.
- Mousavi SH, Moallem SA, Mehri S, Shahsavand Sh, Nassirli H, Malaekhe-Nikouei B. 2011. Improvement of cytotoxic and apoptogenic properties of crocin in cancer cell lines by its nanoliposomal form. *Pharm biol*, 49:1039–1045.
- Neamati N, Boskabady MH. 2010. Effect of *Crocus sativus* (saffron) on muscarinic receptors of guinea pig tracheal chains. *Func Plant Sci Biotec*, 4:128–131.
- Nemati H, Boskabady MH, Ahmadzadeh Vostakolaei H. 2008. Stimulatory effect of *Crocus sativus* (saffron) on  $\beta$  2-adrenoceptors of guinea pig tracheal chains. *Phytomedicine*, 15: 1038–1045.
- Pitsikas N, Zisopoulou S, Tarantilis P A, Kanakis Ch D, Polissiou M G, Sakellaridis N. 2007. Effects of the active constituents of *Crocus sativus* L., crocins on recognition and spatial rats' memory. *Behav brain res*, 183: 141–146.
- Premkumar K, Abraham SK, Santhiya ST, Gopinath PM, Ramesh A. 2001. Inhibition of genotoxicity by saffron (*Crocus sativus* L.) in mice. *Drug chem toxicol*, 24; 421–428.
- Razavi M, Hosseinzadeh H, Abnous Kh, Motamedshariaty V, Imenshahidi M. 2013. Crocin restores hypotensive effect of

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- subchronic administration of diazinon in rats. Iran J Basic Med Sci. 16: 64.
- Razavi Marjan, Hosseinzadeh Hossein, Abnous Khalil, Motamedshariaty Vahideh Sadat, Imenshahidi Mohsen. Crocin Restores Hypotensive Effect of Subchronic Administration of Diazinon in Rats. Iran J Basic Med Sci, Vol. 15, Article in press
- Revelta MP, Cantabrana B, Hidalgo A. 2000. Mechanisms involved in kaempferol-induced relaxation in rat uterine smooth muscle. Life Sci, 67:251-9.
- Rios JL, Recio MC, Giner RM, Manes S. 1996. An update review of saffron and its active constituents. Phytother Res, 10: 189–193.
- Soeda S, Ochiai T, Shimeno H, Saito H, Abe K, Tanaka H, Shoyama Y . 2007. Pharmacological activities of crocin in saffron. J Nat Med, 61: 102–111.
- Srivastava R, Ahmed H, Dixit RK, Dharamveer, Saraf SA. 2010. *Crocus sativus* L. A comprehensive review. Pharmacogn Rev, 4: 200.
- Tarantilis PA, Polissiou MG. 1997. Isolation and identification of the aroma components from saffron (*Crocus sativus*). J Agric Food Chem, 45: 459–462.
- Tang FT, Qian ZY, Liu PQ, Zheng SG, He SY, Bao LP, Huang HQ. 2006. Crocetin improves endothelium-dependent relaxation of thoracic aorta in hypercholesterolemic rabbit by increasing eNOS activity. Biochem Pharmacol, 72:558-565.
- Williams BA, Liu C, De Young L, Brock GB, Sims SM. 2005. Regulation of intracellular  $Ca^{2+}$  release in corpus cavernosum smooth muscle: synergism between nitric oxide and cGMP. Am J Physiol Cell Physiol , 288: C650–C658.
- Winterhalter P, Straubinger M. 2000. Saffron-renewed interest in an ancient spice. Food Rev Int, 16:39–59.
- Xi L, Qian Z, Du P, Fu J. 2007. Pharmacokinetic properties of crocin (crocetin digentiobiose ester) following oral administration in rats. Phytomedicine, 14: 633–636.
- Xu GL, Qian ZY, Yu, SQ, Gong, ZN, Shen, XC. 2006. Evidence of crocin against endothelial injury induced by hydrogen peroxide in vitro. J Asian Nat Prod Res, 8:79–85.
- Xu YC, Leung SWS, Leung GPH, Man RYK. Kaempferol enhances endothelium-dependent relaxation in the porcine coronary artery through activation of large-conductance  $Ca(2+)$ -activated  $K(+)$  channels. Br J Pharmacol 2015 Feb 5. Epub 2015 Feb 5.
- Zhang R, Qian ZY, Han XY, Chen Z, Yan JL, Hamid A. 2009. Comparison of the effects of crocetin and crocin on myocardial injury in rats. Chinese J Nat Med, 7: 223–227.