

Original Research Article

Protective effects of *Elaeagnus angustifolia* L. fruit extract on CCl₄-induced oxidative stress and inflammation in rats liver

Masoud Ojarudi^{1,2}, Ali Golchin^{3,4}, Hamid Reza Karamdel², Mohammad Valilo², Parviz Ranjbarvan^{4,5,*}

¹Student Research Committee, Urmia University of Medical Sciences, Urmia, Iran

²Department of Biochemistry, Faculty of Medicine, Urmia University of Medical Sciences, Urmia, Iran

³Solid Tumor Research Center, Cellular and Molecular Medicine Research Institute, Urmia University of Medical Sciences, Urmia, Iran

⁴Department of Applied Cell Sciences, School of Medicine, Urmia University of Medical Sciences, Urmia, Islamic Republic of Iran

⁵Cellular and Molecular Research Center, Cellular and Molecular Medicine Research Institute, Urmia University of Medical Sciences, Urmia, Iran

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* Corresponding Author:

Tel: 09143061276

Fax: 04432240642

ranjbarvan@gmail.com

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Abstract

Objective: Hepatic cells face oxidative stress-induced damage, but plant antioxidants may offer protection. This study aimed to assess *Elaeagnus angustifolia* L. fruit extract's potential in shielding rat livers from CCl₄ damage.

Materials and Methods: 30 Male Wistar rats were randomly divided into five groups: normal control (received distilled water), *E. angustifolia* hydroalcoholic extract control, CCl₄ control, *E. angustifolia* extract pretreatment (600 mg/kg), and silymarin pretreatment (100 mg/kg). After 14 days of oral administration of extracts, CCl₄ was injected intraperitoneally. The samples were collected 48 hr later. Histological and biochemical analyses were then carried out.

Results: CCl₄ injection caused significant ($p < 0.001$) changes in liver serum enzymes, lipid profile, bilirubin, total protein, serum albumin, antioxidant enzymes, malondialdehyde, Inflammatory cytokines, and liver tissue morphology. *E. angustifolia* extract pretreatment significantly ($p < 0.05$) returned changes to the normal state.

Conclusion: This study's findings revealed that *E. angustifolia* extract pretreatment could reduce liver injury caused by CCl₄ in rats.

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Introduction

The liver is crucial for processing exogenous and endogenous compounds, making it susceptible to daily oxidative stress (Ahmadi *et al.* 2022; Yu *et al.* 2018). Excessive ROS (Reactive oxygen species) from various sources can damage nucleic acids, proteins, lipids, and membranes, affecting liver function despite antioxidant defenses (Ullah *et al.* 2020). Hence, investigating this field is imperative, and animal models can be utilized for studying liver damage induced by oxidative stress in research.

CCl₄ is a known inducer of oxidative liver damage in laboratory animals. CCl₄ induces oxidative liver damage by forming radicals that bind to biomolecules, causing lipid peroxidation and membrane damage, exacerbating inflammation and injury (Rezagholizadeh *et al.* 2022).

Exploring natural sources for antioxidant compounds can be a promising approach to prevent oxidative damage. Medicinal plants provide antioxidants like polyphenols and flavonoids for protection (El-Yagoubi *et al.* 2024; Valvi *et al.* 2016). *Elaeagnus angustifolia* L. fruit is also rich in antioxidant compounds.

Elaeagnus angustifolia L., a small tree or shrub, is rich in phytochemicals like terpenoids, flavonoids, tannins, and vitamins. Main flavonoids include catechin, epigallocatechin, and chlorogenic acid (Hassanzadeh and Hassanpour 2018).

Silymarin, derived from *Silybum marianum* and commonly referred to as milk thistle, is an extract that has been utilized for centuries in the treatment of liver diseases (Gillissen and Schmidt 2020). The flavonoid silymarin, along with its structural component silibinin, are compounds that have been shown to possess hepatoprotective properties (Fraschini *et al.* 2002). In most studies, silymarin is employed as a standard treatment for liver conditions (Barreiro Carpio *et al.* 2024; El Rabey *et al.* 2021).

Hence, based on the antioxidant properties found in the fruit of this plant,

this study aimed to evaluate hepatoprotective effects of *E. angustifolia* L. fruits in the CCl₄-induced rats.

Materials and Methods

Chemicals

Chemicals were sourced from Merck (Germany), including methanol, thiobarbituric acid, serum albumin, hydrogen peroxide, and more. Reagents like Folin-Ciocalteu, DPPH (2,2-diphenyl-1-picrylhydrazyl), gallic acid, and quercetin were from Sigma. Serum parameters were analyzed using kits from Pars-Peyvand Co., while SOD (Superoxide dismutase), GPx (Glutathione peroxidase), TNF- α (Tumor necrosis factor alpha), and IL-6 (Interleukin 6) assay kits were from Zell Bio and karmania pars gene Co, Kerman, Iran.

Extraction method

Dried fruit of *E. angustifolia* was purchased from Iran Extract Research Institute (The genus and species were definitively identified by a botanist), ground, and macerated in 70% methanol (500 g of *E. angustifolia* fruit powders in 2 L of 70% methanol) for four days. The resulting macerate was filtered, dried at room temperature, freeze-dried for two days, and stored in a freezer (the maceration process yielded 80 g of crude *E. angustifolia* extract) (Ojarudi *et al.* 2020).

Total phenolic, total flavonoid content, and DPPH radical scavenging activity

Total phenolic content was assessed using the Folin-Ciocalteu method, while total flavonoid content was determined through a colorimetric method similar to Kadhim *et al.* (Jawad Kadhim *et al.* 2019). Antioxidant activity was measured using the DPPH radical scavenging method (Csicsor and Tombácz 2022).

Animals and experimental procedure

The study utilized 30 male Wistar rats (200-220 g) housed in a temperature-controlled room with a 12-hour light-dark

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cycle. Animals were divided onto five different groups at random:

1- Normal Control: received distilled water orally + olive oil intraperitoneally

2- *E. angustifolia*: received *E. angustifolia* fruit extract orally (600 mg/kg) + olive oil intraperitoneally

3- CCl₄: received distilled water orally + CCl₄ intraperitoneally

4- *E. angustifolia* + CCl₄: received *E. angustifolia* fruit extract orally (600 mg/kg) + CCl₄ intraperitoneally

5- Silymarin + CCl₄: received Silymarin orally (100 mg/kg (Ojarudi et al. 2020)) + CCl₄ intraperitoneally

Considering the previous studies indicating that the optimal therapeutic dosage of *E. angustifolia* fruit extract is 600 mg/kg, this dosage was used in the current research (Dabbaghmanesh et al. 2017). Also, according to similar studies (Ojarudi et al. 2020), the pretreatment group receiving silymarin was used to compare the pretreatment effects of *E. angustifolia* extract.

All groups were treated for 14 days. On day 14, groups three, four, and five received 1 ml/kg of a CCl₄ and olive oil mixture intraperitoneally, while groups one and two received only olive oil (the solvent of CCl₄) (Mahmoodzadeh et al. 2017). After 48 hr and a 12-hr fast, animals were anesthetized (Intraperitoneally injection of 10 mg/kg ketamine and 90 mg/kg xylazine per body weight), blood and liver tissue samples were collected for analysis.

Serum factor levels

Serum factors, including liver enzymes (AST, ALT, ALP, and GGT [Gamma glutamyl transferase]), total protein, albumin, lipid profile (total cholesterol, triglycerides, LDL-C, and HDL-C), and bilirubin, were analyzed using an autoanalyzer (Biochemistry Analyzer BT 3000, Italy) following the instructions provided in each laboratory kit.

Protein assay

Tissue protein was measured using Bradford's method (Bradford 1976).

MDA

The concentration of liver tissue MDA was determined using the Uchiyama & Mihara technique, with little modification similar to our earlier work (Mihara and Uchiyama 1978; Ojarudi et al. 2020).

Total antioxidant capacity

The FRAP method was used to determine total antioxidant capacity (Benzie and Strain 1996).

Antioxidant enzymes

The enzymatic activity of CAT (Catalase) present in the tissue was quantified utilizing the Aebi's method (Aebi 1984). Furthermore, the activity of both SOD and GPx enzymes was evaluated using the Zell Bio kit's procedure.

Inflammatory cytokines measurement

The levels of TNF- α and IL-6 were measured by the relevant commercial ELISA kit with the manufacturer's protocol.

Histopathological study

Liver tissue samples were fixed in 10% formalin, dehydrated in ethanol (50-100%), cleared with Xylene, embedded in paraffin, sectioned (4-5 μ m), and stained with hematoxylin-eosin.

Liver index and weight

The body mass and liver mass of each rat were measured at the conclusion of the experiment. The liver index (Eidi et al. 2013b), which is the ratio of liver mass to body mass, was then computed using the following formula:

Liver index = (liver weight/ body weight) \times 100%

Statistical analysis

Data is reported as mean \pm standard deviation. Data analysis was done by one-way analysis of variance with GraphPad Prism 8.3.0. Post hoc comparisons were

made using Tukey's test to determine specific group differences. A significance level of $p < 0.05$ was considered. The experiments were conducted in triplicate to ensure the reliability and reproducibility of the results.

Results

Contents of total phenolic and total flavonoid and antioxidant activity

Table 1 displays the phenolic and flavonoid content as well as the *in vitro* antioxidant activity of the extract.

Table 1. Phenolic and flavonoid content and antioxidant activity of the *Elaeagnus angustifolia* L. fruit extract

Sample	Total phenolic content (mg GA/gram of extract)	Total flavonoid content (mg QU/gram of extract)	DPPH scavenging activity IC ₅₀ value (µg/ml)
<i>E. angustifolia</i>	36.04±0.49	23.81±0.71	144.71

GA: Gallic acid, QU: Quercetin

Table 2. The effect of *Elaeagnus angustifolia* L. fruit extract on liver enzymes

Groups	AST (IU/L)	ALT (IU/L)	ALP (IU/L)	GGT (IU/L)
Normal Control	103.16±4.49	48.83±3.65	429.16±6.55	1.42±0.29
<i>E. angustifolia</i>	102.33±3.61	48.66±2.58	423.66±10.38	1.48±0.17
CCl ₄	709.50±12.56 ^a	631.33±14.76 ^a	713.50±34.58 ^a	6.21±0.76 ^a
<i>E. angustifolia</i> + CCl ₄	303.66±7.96 ^d	253.83±13.51 ^d	511.50±10.98 ^d	4.14±0.63 ^c
Silymarin + CCl ₄	329.66±10.30 ^d	252.50±15.52 ^d	508.83±12.87 ^d	4.23±0.69 ^c

Data is expressed as the mean ± SD. ^a $p < 0.001$ compared with normal control; ^b $p < 0.05$, ^c $p < 0.01$, and ^d $p < 0.001$ compared with CCl₄

Effects of *E. angustifolia* extract administration on bilirubin, protein and serum albumin

CCl₄ injection significantly elevated total and direct bilirubin levels ($p < 0.001$) and decreased total protein and serum albumin levels ($p < 0.001$). Pretreatment with *E. angustifolia* extract mitigated these effects ($p < 0.05$), as shown in Table 3.

Effects of *E. angustifolia* extract administration on lipid profile

CCl₄ administration increased total cholesterol, triglycerides, and LDL-C levels, while decreasing HDL-C levels ($p < 0.001$). Pretreatment with *E. angustifolia* extract attenuated these changes ($p < 0.01$), as shown in Table 4.

Antioxidant activity is presented as 50% inhibition of free radicals (IC₅₀), representing the extract concentration (µg/ml) needed to scavenge 50% of DPPH radicals.

Effects of *E. angustifolia* extract administration on liver enzymes

CCl₄ administration significantly increased liver enzyme levels (AST, ALT, ALP, and GGT) ($p < 0.001$), which were attenuated by pretreatment with *E. angustifolia* extract ($p < 0.01$), as shown in Table 2.

Effects of *E. angustifolia* extract administration on lipid peroxidation

Figure 1 shows that CCl₄ administration significantly increased MDA levels compared to the normal control group ($p < 0.001$). Pretreatment with *E. angustifolia* extract reduced MDA levels significantly ($p < 0.001$).

Effects of *E. angustifolia* extract administration on total antioxidant capacity

As shown in Figure 2, CCl₄ significantly decreased total antioxidant capacity ($p < 0.001$). Pretreatment with *E. angustifolia* extract reversed this decline, significantly increasing total antioxidant capacity in liver tissue samples ($p < 0.01$).

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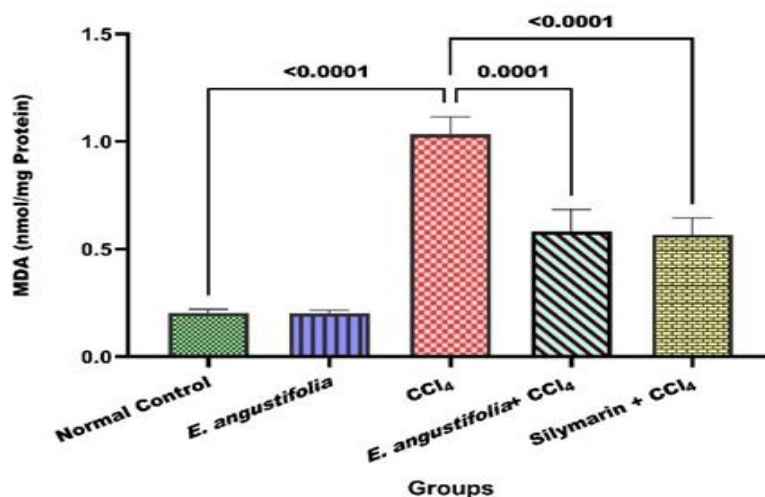


Figure 1. The effect of *Elaeagnus angustifolia* L. fruit extract on the level of lipid peroxidation. Data is expressed as the mean \pm SD.

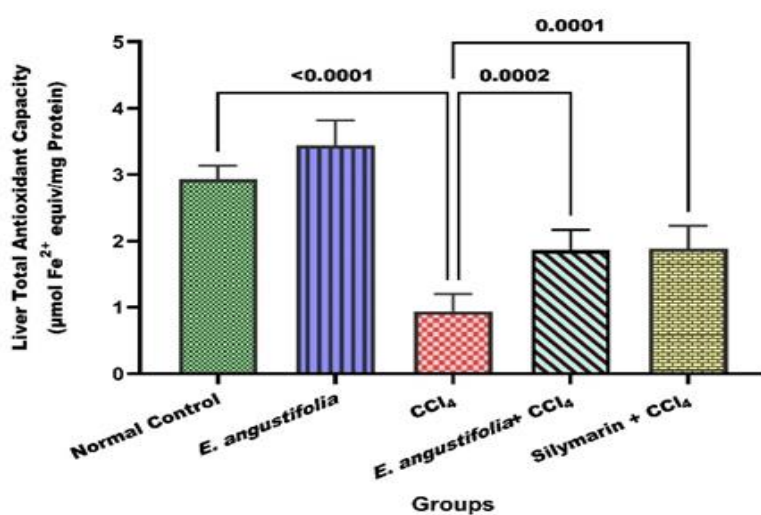


Figure 2. The effect of *Elaeagnus angustifolia* L. fruit extract on total antioxidant capacity. Data is expressed as the mean \pm SD.

Table 3. The effect of *Elaeagnus angustifolia* L. fruit extract on serum bilirubin, total protein, and albumin levels

Groups	BilT (mg/dl)	BilD (mg/dl)	Total Protein (g/dl)	Albumin (g/dl)
Normal Control	0.65 \pm 0.03	0.08 \pm 0.01	7.19 \pm 0.13	3.18 \pm 0.17
<i>E. angustifolia</i>	0.64 \pm 0.02	0.07 \pm 0.01	7.12 \pm 0.44	3.23 \pm 0.19
CCl ₄	1.11 \pm 0.11 ^a	0.32 \pm 0.03 ^a	5.03 \pm 0.48 ^a	2.36 \pm 0.24 ^a
<i>E. angustifolia</i> + CCl ₄	0.84 \pm 0.08 ^b	0.21 \pm 0.03 ^d	6.21 \pm 0.55 ^c	2.91 \pm 0.11 ^d
Silymarin + CCl ₄	0.87 \pm 0.08 ^b	0.21 \pm 0.02 ^d	6.07 \pm 0.68 ^c	2.88 \pm 0.11 ^d

Data is expressed as the mean \pm SD. ^ap<0.001 compared with normal control; ^bp<0.05, ^cp<0.01, and ^dp<0.001 compared with CCl₄

Table 4. Effect of *Elaeagnus angustifolia* L. fruit extract on lipid profile

Groups	Cholesterol (mg/dl)	Triglyceride (mg/dl)	LDL (mg/dl)	HDL (mg/dl)
Normal Control	68.66±3.50	48.38±2.77	26.50±1.87	39.41±1.50
<i>E. angustifolia</i>	69.16±2.63	46.51±3.81	24.83±2.63	38.81±2.19
CCl ₄	87.16±6.33 ^a	191.51±6.58 ^a	51.83±5.81 ^a	24.96±1.92 ^a
<i>E. angustifolia</i> + CCl ₄	74.50±4.27 ^d	142.50±4.84 ^d	34.50±4.13 ^d	31.83±2.74 ^d
Silymarin + CCl ₄	72.83±5.63 ^d	131.33±5.98 ^d	33.66±4.08 ^d	30.53±2.63 ^c

Data is expressed as the mean ± SD. ^ap<0.001 compared with normal control; ^bp<0.05, ^cp<0.01, and ^dp<0.001 compared with CCl₄

Effects of *E. angustifolia* extract administration on antioxidant enzymes

A single CCl₄ injection significantly reduced antioxidant enzyme (CAT, SOD, and GPx) activity (p<0.001). Pretreatment with *E. angustifolia* extract prevented this reduction significantly (p<0.05), as shown in Figure 3.

Effects of *E. angustifolia* extract administration on inflammatory cytokines

Serum TNF- α and IL-6 significantly increased with CCl₄ injection (p<0.001). Pretreatment with *E. angustifolia* extract significantly prevented this increase, similar to silymarin (Figure 4).

Histopathological observations (Figure 5 and Table 5) confirmed the protective effect of *E. angustifolia* fruit extract against liver damage. CCl₄ induced extensive changes in the lobules, such as fat accumulation, cellular vacuolation and necrosis, sinusoidal dilation and inflammatory cell infiltration, in the CCl₄ group. However, the extract preserved hepatocyte structure and reduced necrosis and inflammation, similar to silymarin.

Liver index

The CCl₄-injured group showed a significant increase (p<0.05) in liver index. Pretreatment with *E. angustifolia* extract led to a significant decrease (p<0.05) in this index (Table 6).

Histopathological study

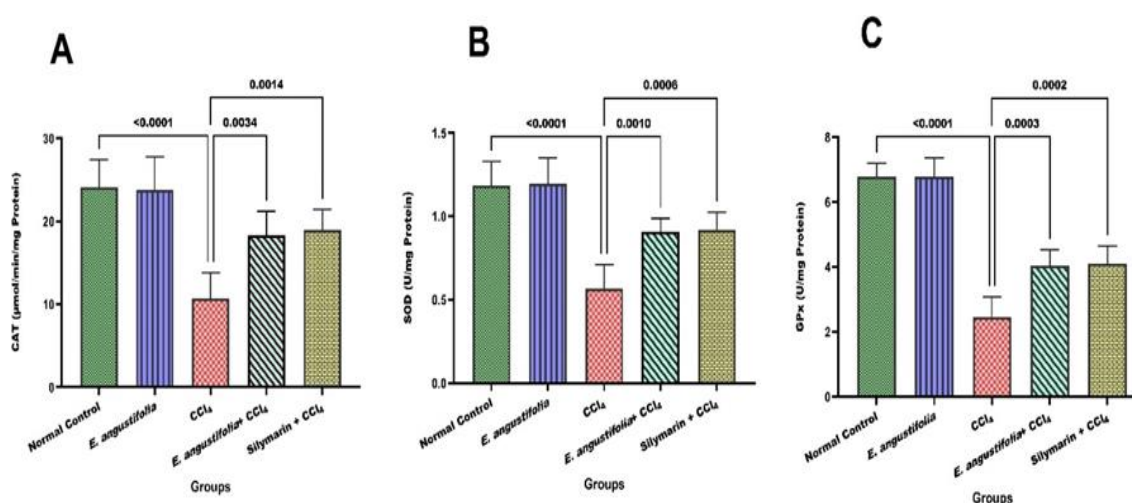


Figure 3. The effect of *Elaeagnus angustifolia* fruit extract on antioxidant enzymes activity. Data is expressed as the mean ± SD. (A: Catalase, B: SOD, and C: GPx)

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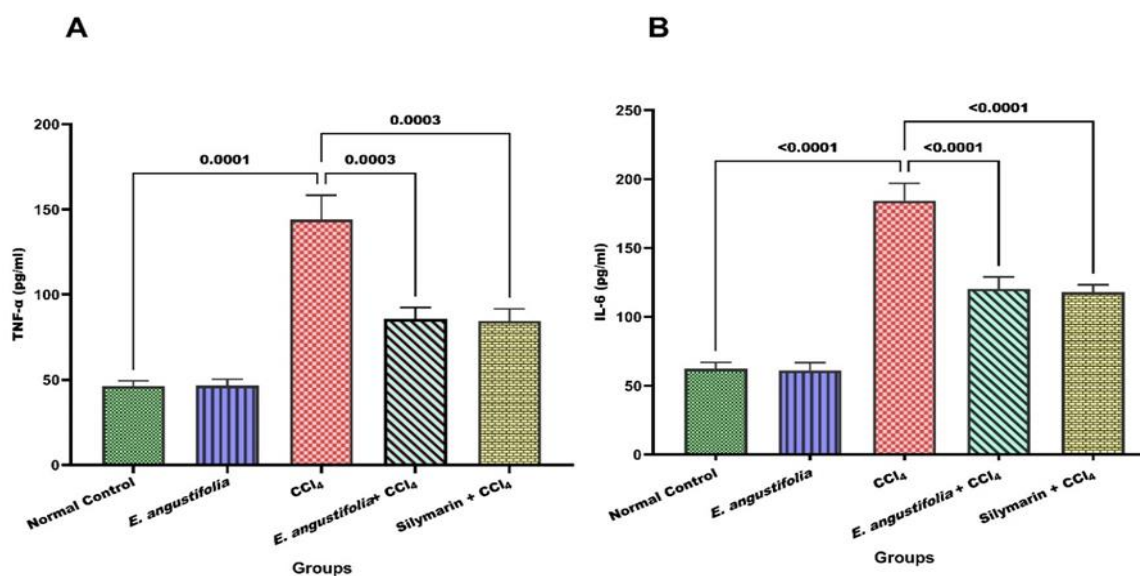


Figure 4. The effect of *Elaeagnus angustifolia* L. fruit extract on inflammatory cytokines. Data is expressed as the mean \pm SD. (A: *TNF- α* and B: *IL-6*)

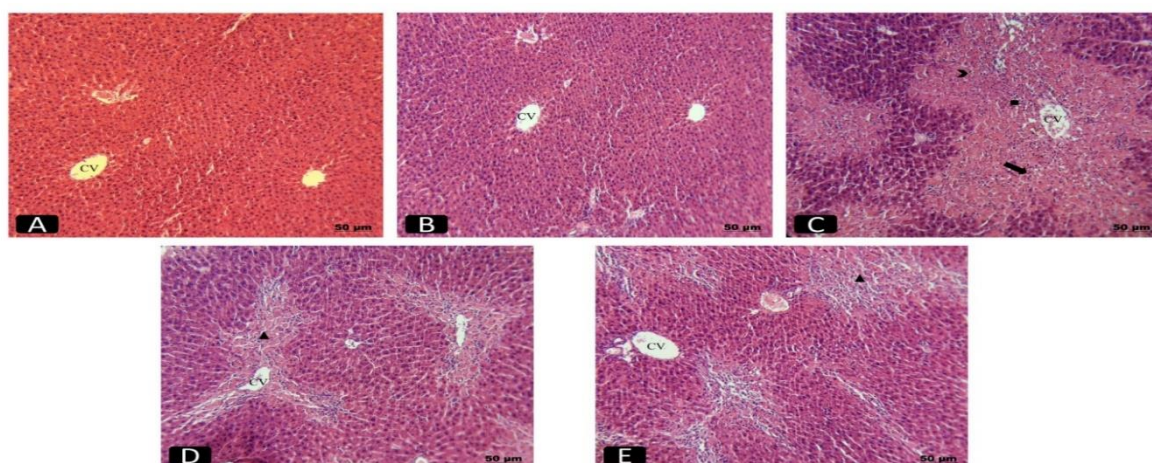


Figure 5. Microscopic images of liver tissue in different groups: A- Normal control group. B- *E. angustifolia* group. C- CCl₄ group. D- *E. angustifolia* + CCl₄ group. E- Silymarin + CCl₄ group. CV Central vein. ■ Necrosis. ▴ Irregular cell plates. → Inflammation. ▲ Hepatocyte degeneration

Table 5. Histological injury score of the liver

Groups	Index							
	Injury of Score ^a							
	Hepatocyte degeneration	Sinusoidal dilation	Portal area dilation	Irregular cell plates	Necrosis	Inflammation	Fatty degeneration	Total scores
Normal control	0	0	0	0	0	0	0	0
<i>E. angustifolia</i>	0	0	0	0	0	0	0	0
CCl ₄	3	3	2	3	3	2	2	18
<i>E. angustifolia</i> + CCl ₄	2	1	1	2	2	1	1	10
Silymarin + CCl ₄	1	1	1	1	1	1	1	7

^a The degree of hepatic damage was assessed by examining the livers under a microscope and assigning a score based on the following criteria: score 0=no signs of cell injury; 1= hepatocyte injury on less than 25% of the tissue; score 2= hepatocyte injury on 25–50% of the tissue; score 3= immense, but focal, hepatocyte injury; score 4= overall hepatocyte necrosis.

Table 6. The effect of *Elaeagnus angustifolia* L. fruit extract on liver index

Groups	Body weight on day	Body weight on day	Liver weight	Liver index
	1 (gram)	16 (gram)	(gram)	(%)
Normal control	207.66±5.00	247.33±5.68	7.26±0.25	2.94±0.15
<i>E. angustifolia</i>	209.16±5.41	249.16±3.06	7.43±0.36	2.98±0.14
CCl ₄	210.33±6.18	243.66±6.08	10.90±0.46 ^a	4.64±0.18 ^a
<i>E. angustifolia</i> + CCl ₄	209.66±6.43	243.33±7.31	8.78±0.30 ^d	3.61±0.09 ^d
Silymarin + CCl ₄	208.16±6.79	242.83±5.77	8.63±0.36 ^d	3.55±0.18 ^d

Data is expressed as the mean ± SD. ^a p<0.001 compared with normal control; ^b p<0.05, ^c p<0.01, and ^d p<0.001 compared with CCl₄

Discussion

This study evaluated *E. angustifolia* fruit extract for antioxidant and anti-inflammatory effects against CCl₄-induced liver injury, demonstrating efficacy in reducing damage. Biochemical markers were used to assess oxidative damage.

The cell membrane, rich in fatty acids, is highly susceptible to free radical damage. Leakage of cell contents into the bloodstream indicates cellular and tissue damage severity, assessable by measuring specific molecules in the blood (Stark 2005). Assessing liver enzymes in serum is a common method to evaluate liver damage (Meng *et al.* 2020). Significantly (p<0.001) elevated levels of ALT, AST, ALP, and GGT indicate hepatocyte leakage due to CCl₄-induced liver injury in rats compared to the normal group. Liver damage also led to decreased serum protein and albumin levels, altered lipid profiles, and changes in bilirubin levels similar to the findings of other researchers (Ebeid *et al.* 2015; Saleh Gazwi and Mahmoud 2019; Ullah *et al.* 2020). Pretreatment with *E. angustifolia* fruit extract, rich in phenolic and flavonoid content, remarkably (p<0.01) protected against liver damage, potentially through its antioxidant properties, and via neutralizing free radicals and preventing further hepatocyte damage. We also investigated the antioxidant markers in liver tissue to further elucidate the liver-protection effects of *E. angustifolia* fruit extract.

MDA, a marker of lipid peroxidation, was utilized to evaluate oxidative damage in the liver (Gawel *et al.* 2004). CCl₄-induced free radicals can induce lipid peroxidation, causing breakdown of PUFA (Polyunsaturated fatty acids) molecules and harmful compound production (Negre-Salvayre *et al.* 2010). Studies have demonstrated that CCl₄ raises MDA levels in rat liver tissue (Chen *et al.* 2020; Eltahir *et al.* 2020). CCl₄ notably (p<0.001) increased lipid peroxidation, but *E. angustifolia* fruit extract significantly (p<0.01) decreased MDA levels, suggesting protection against oxidative damage.

Further investigation should focus on the impact of *E. angustifolia* fruit extract on the antioxidant enzyme system in liver tissue. Cellular antioxidant enzymes neutralize radicals and protect cells. However, the efficacy of this protective system diminishes when the balance of antioxidants and free radicals within the cell is disrupted (Mohammadi Mahjoob *et al.* 2024; Tahavvori *et al.* 2023; Wei *et al.* 2021). Exposure to CCl₄ is known to disrupt this system, leading to cellular dysfunction (Almatroodi *et al.* 2020). CCl₄ caused a significant (p<0.001) decrease in antioxidant enzymes (CAT, SOD, and GPx) and pretreatment with the extract preserved enzyme activity, indicating its potential to enhance antioxidant defense against free radicals.

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CCl₄-induced liver damage also involves inflammation as characterized by increased TNF- α and IL-6 levels (Long et al. 2022). This study confirmed elevated cytokines due to inflammation, consistent with previous research (Jaime-Pérez et al. 2020; Long et al. 2022; Said et al. 2022). The *E. angustifolia* fruit extract was found to inhibit the elevation of TNF- α and IL-6 levels, indicating the anti-inflammatory properties of the extract's compounds.

Histological analysis of rat liver tissue confirmed CCl₄-induced damage, showing necrosis, cell infiltration, fibrosis, and steatosis like previous studies (Eidi et al. 2013a). Pre-treatment with *E. angustifolia* fruit extract improved liver structure, consistent with biochemical results, suggesting its potential for liver protection and tissue regeneration. Multiple studies have confirmed the extract's ability to reduce inflammation. This is achieved through the inhibition of cyclooxygenase, highlighting its potential therapeutic benefits in inflammatory conditions (Hamidpour et al. 2017).

Finally, the liver index, an important marker for assessing the severity of liver damage, was utilized to demonstrate the protective effects of the extract (Wei et al. 2021). CCl₄ significantly elevated the liver index, consistent with previous research. Pre-treatment with the extract effectively ($p < 0.01$) mitigated this increase, similar to silymarin. These results underscore the potential of *E. angustifolia* fruit extract's antioxidant properties in protecting against severe liver damage induced by CCl₄.

This study revealed that *E. angustifolia* fruit extract exhibits antioxidant effects and enhances liver function against CCl₄-induced liver injury in rats.

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Conflicts of interest

The authors report no conflicts of interest.

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Ethical Considerations

Ethical approval for this study was obtained from the Urmia university of medical sciences Animal Ethics Committee (IR.UMSU.AEC.1402.003).

Code of Ethics

IR.UMSU.AEC.1402.003

Authors' Contributions

PR oversaw the study design and supervised its execution, as well as revising the final manuscript. MO, MV, and HRK carried out the experimental procedures. AG conducted the statistical analyses. MO wrote the initial draft of the manuscript. All authors have read and approved the final manuscript.

Abbreviations

ALP	Alkaline phosphatase
ALT	Alanine Transaminase
AST	Aspartate aminotransferase
CAT	Catalase
CCl ₄	Carbon tetrachloride
DPPH	2,2-diphenyl-1-picrylhydrazyl
FRAP	Ferric Reducing Antioxidant Power Assay
GGT	Gamma glutamyl transferase
GPx	Glutathione peroxidase
HDL-C	High Density Lipoprotein Cholesterol
IC50	Half-maximal inhibitory concentration
IL-6	Interleukin 6
LDL-C	Low Density Lipoprotein Cholesterol
MDA	Malondialdehyde
PUFA	Polyunsaturated fatty acids
ROS	Reactive oxygen species

SOD Superoxide dismutase
 TNF- α Tumor necrosis factor alpha

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