

Review article

Antioxidant activities of kombucha tea: a systematic review and meta-analysis of *in vitro* studies

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Abstract

Objective: Human body is always exposed to oxidants which could be suppressed with antioxidant supplementation. Kombucha is a fermented tea beverage that has been shown to have antioxidant properties. The current systematic review and meta-analysis aimed to evaluate the antioxidant potential of kombucha tea based on the existing literature.

Materials and methods: A systematic literature search following the PRISMA 2020 guidelines was conducted to include eligible studies (original research in English language on the antioxidant activity of kombucha tea, with available full text). Data extraction was performed independently by three reviewers. Overall, the study process was undertaken from September 2024 to May 2025; final search conducted on September 11, 2024. Random-effects meta-analysis was selected to summarize the statistical information of the antioxidative-related parameters.

Results: Fourteen studies were included in this systematic review and meta-analysis. Our results showed that kombucha had a significant effect against unfermented tea control in 2,2-diphenyl-1-picrylhydrazyl/DPPH assays (% radical scavenging ability = 9.84% (95% CI = 4.63 to 15.05%, $p = 0.0002$)) and half-maximal inhibitory concentration (IC_{50}) = -64.54 mg/L (95% CI = -99.06 to -30.03 mg/L, $p = 0.0002$). However, other parameters such as 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)/ABTS assays, ferric reducing antioxidant power, intracellular reactive oxygen species (ROS), total phenolic compounds, and total flavonoid contents did not differ significantly between kombucha and control groups.

Conclusion: The current systematic review and meta-analysis presented a significant difference of antioxidant capacity (DPPH) of kombucha tea compared to control (unfermented tea). Nevertheless, due to high heterogeneity in the included studies, more studies should be conducted to validate kombucha tea antioxidant activity.

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Introduction

Antioxidants are defined as any substance with the ability to neutralize reactive oxygen species (ROS) and their derivatives, such as reactive nitrogen species (RNS) or reactive sulphur species (RSS) (Martemucci et al. 2022; Baruah et al. 2025). ROS are the by-products of cellular metabolism formed following mitochondrial oxidases and other cellular compartments activity (Gulcin 2025). Meanwhile, RNS and RSS are formed through ROS reaction with nitric oxide and thiol, respectively (Salehi et al. 2018; Mandal et al. 2022).

Although ROS are naturally produced in our bodies, their overproduction may occur in some situations such as aging, psychological stress, infection, inflammation, cancer, and excessive exercise (Pizzino et al. 2017; Jena et al. 2023). ROS elevation could induce cellular damage via senescence induction and increased production of inflammatory cytokines. Antioxidants are vital to prevent the ROS-associated cellular damage (Salehi et al. 2018; Kozlov et al. 2024). Furthermore, antioxidants are also beneficial to prevent the development of various chronic diseases including cancer, cardiovascular disease, and inflammatory or autoimmune disease (Zehiroglu and Ozturk Sarikaya 2019; Ayoka et al. 2022; Blagov et al. 2024). Herbal substances may be beneficial to help in improving antioxidative activities and preserve human health (Parham et al. 2020; Stevanny et al. 2020; Sukweenadhi et al. 2020).

Kombucha is a traditional beverage that originated from Manchuria, China and traditionally produced through the fermentation of tea leaves (*Camellia sinensis*) (Barros et al. 2024; Falasifah et al. 2025). It involved a symbiotic culture of bacteria and yeast (SCOBY) consisting of acetic acid bacteria (*Komagataeibacter*, *Gluconobacter*, and *Acetobacter*), lactic acid bacteria (*Lactobacillus* and *Lactococcus*), and osmophilic yeast (*Schizosaccharomyces pombe*,

Saccharomycodes ludwigii, *Kloeckera apiculata*, *Saccharomyces cerevisiae*, *Zygosaccharomyces bailii*, *Torulaspora delbrueckii*, and *Brettanomyces bruxellensis*). This fermentation process occurs in an aerobic and high in sucrose (5–8% sugar) medium for several days (Villarreal-Soto et al. 2018; de Miranda et al. 2022; Anantachoke et al. 2023; Sarshar et al. 2025). Although several studies have reported the antioxidant properties of kombucha and its potential health benefits (Hananta et al. 2025), the results have been controversial. Jakubczyk et al (2020) have reported best antioxidant properties of green tea kombucha compared with black, white, and red tea. Meanwhile, Cardoso et al (2020) reported a higher antioxidant capacity in black tea kombucha than green tea kombucha, although green tea types exhibited higher antibacterial and antiproliferative activities. On the other hand, a study reported that green tea exhibited greater antioxidant potential than kombucha with no difference in the concentration of total phenolic compounds (TPCs) (Teixeira Oliveira et al. 2023). Consequently, there is no definitive agreement regarding the antioxidant impact of kombucha in comparison to its unfermented substrate. To our knowledge, no systematic review has yet offered a quantitative synthesis (meta-analysis) of these data. This gap in the literature makes it challenging to determine whether the fermentation process increases the antioxidant potential *in vitro*. Therefore, we conducted a systematic review and meta-analysis aimed to summarize the findings of previous studies regarding antioxidant activity of kombucha tea.

Materials and Methods

This review followed the Preferred Reporting Items guidelines for Systematic Reviews and also Meta-analysis (PRISMA) 2020 (Page et al. 2021). Since the current systematic review included *in vitro* studies, the protocol could not be published in the

international prospective register of systematic reviews (PROSPERO). Supplementary Table 1 shows the PRISMA 2020 checklist filled following the completion of this manuscript writing.

Literature search

A comprehensive literature search was conducted to assess the antioxidant activity of kombucha tea from seven databases: PubMed, ProQuest, Taylor and Francis, SAGE, JSTOR, Wiley, and Emerald. The

review was conducted between September 2024 and May 2025. The articles were retrieved with no publication date and geographical restriction. The search terms applied in the current systematic review are listed in Table 1. Literature search was limited to those published in English language and with available full-text. In addition, we manually searched additional possible studies from the references of the included studies.

Table 1. Search terms

Databases	Search terms
PubMed	((("reactive Oxygen Species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")
ProQuest	("reactive oxygen species") AND ("antioxidant") AND ("kombucha")
Taylor and Francis	((("reactive oxygen species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")
Sage	((("reactive oxygen species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")
JSTOR	((("reactive oxygen species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")
Wiley	((("reactive oxygen species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")
Emerald	((("reactive oxygen species") OR ("ROS")) AND ("antioxidant")) AND ("kombucha")

Study selection

The inclusion criteria are: (1) original article, (2) written in English, (3) accessible full-text, and (4) investigated the antioxidant activity of kombucha tea. Meanwhile, the exclusion criteria are: (1) secondary article sources including review, editorial, correspondence, letter, or commentary, (2) no appropriate control (unfermented tea), or (3) no antioxidant markers detection. The current review primarily included studies which reported *in vitro* antioxidant assays using chemical-based test. However, to expand the number of included studies, research done using cell-based *in vitro* assay was also included.

Data extraction

Three authors (KT, AMS, and LH) extracted further information independently about the research that has been included, using Rayyan QCRI, a semi-automatic screening tool for systematic review (Ouzzani et al. 2016; Umar et al. 2022, 2023). Inter-rater discrepancies were addressed through meticulous re-evaluation and conversation of the article among evaluators until an accordance was reached,

subsequently verified by a senior author (TPU). The extracted data were as follows: (1) First author and publication year; (2) study design; (3) country of study origin; (4) method; (5) antioxidant activity, including 2,2-diphenyl-1-picrylhydrazyl/DPPH assay, 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)/ABTS assay, ferric ion reducing antioxidant power (FRAP), intracellular ROS generation, TPCs, and total flavonoid contents (TFCs). Missing or incomplete data were estimated from research graphs or diagrams. However, if this was impossible, the studies were excluded.

Quality assessment

The risk of bias analysis was conducted using a tool developed by Silva et al (2020) as an assessment tool for *in vitro* laboratory-based systematic review. The following parameters were considered during the critical appraisal: (i) sample size calculation, (ii) samples with similar dimensions, (iii) control group, (iv) standardization of procedures, (v) statistical analysis, and (vi) other risk of bias. Other risk of bias may include: selective reporting

bias and blinding. Each parameter was determined as “low,” “high,” or “unclear” risk of bias. Three independent reviewers (MMS, SVK, and LL) conducted the assessment, and any discrepancies were resolved through discussion with a senior author (ZA).

Types of outcome measures

The outcome measures assessed in this systematic review included free radical scavenging activity (DPPH-% and half-maximal inhibitory concentration (IC₅₀); ABTS-% and IC₅₀; and FRAP), antioxidative compounds level (TPCs and TFCs), and oxidative stress marker levels (intracellular ROS). All these parameters provided a comprehensive list of different aspects of antioxidant activities.

Statistical analysis

Statistical analyses (meta-analyses) were conducted using RevMan 5.4 (Cochrane Collaboration). The analyzed parameters were numerical variables; thus, it was followed by comparing the previously determined measures the kombucha (green tea leaves are the primary kombucha types tested due to the highest antioxidative capacities (Jakubczyk et al. 2020) and control groups (unfermented tea) to obtain the Standardized Mean Difference (SMD) or Mean Differences (MD) value and 95% Confidence Interval (95% CI). For data presented as a diagram or a graph, the numerical values were extracted using the WebPlotDigitizer version 5.0 (<https://automeris.io/WebPlotDigitizer>; Pacifica, California, USA). The conclusion was declared statistically significant if the overall outcome SMD or MD did not intersect the line 0 with p value < 0.05. Then, heterogeneity analysis was carried out by I² value determination. If the I² value is >50% then, there is significant heterogeneity between studies and random-effect model (REM) will be used.

Otherwise, fixed-effect model (FEM) was used. For variables with significant heterogeneity, sensitivity analyses were undertaken to determine the stability of the results through the leave-one-out meta-analysis

Results

Literature retrieval

Following literature search, 481 articles were retrieved during the preliminary process. After deduplication, 42 duplicate records were removed, leaving 439 articles which undergone abstracts, titles, and keywords screening. The process resulted in 42 relevant papers which were allocated for full-text screening. After full-text screening, 14 studies were included for review and eleven studies were statistically analysed. Figure 1 illustrates the literature retrieval process.

Study characteristics

The characteristics of the included studies are shown in Table 2. All studies (Kaewkod et al. 2019; Fibrianto et al. 2020; Jakubczyk et al. 2020, 2024; Khaleil 2020; Ziemlewska et al. 2021, 2023, 2024; Li et al. 2022; Xu et al. 2022; Zhang et al. 2023; Sales et al. 2023; Xiong et al. 2023; Dechakhamphu et al. 2023) were published between 2019 and 2024. All studies were done as *in vitro* studies (n=14). The studies were conducted in various countries: Poland (n=5), China (n=3), Thailand (n=2), Brazil (n=1), Indonesia (n=1), Saudi Arabia (n=1), and South Korea (n=1). All investigations used tea-based kombucha derived from various types of tea sources. The fermentation period of kombucha ranged from 7 to 30 days. The included studies assessed different antioxidant activity (using ABTS, DPPH, FRAP, TFC, TPC, and intracellular ROS assays).

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Table 2. Characteristics of the included studies

Author (Year)	Country	Methods		Kombucha Antioxidant Activities								
		Tea-Based (exp)	Fermentation duration	Cell objects	DPPH (%)	DPPH (IC ₅₀ , mg/L)	ABTS (%)	ABTS (IC ₅₀ , mg/L)	FRAP (μM Fe(II)/L)	TPC (mg/L)	TFC (mg/L)	Intracellular ROS (%)
Dechakhamphu et al. (2023)	Thailand	<i>Cyperus rotundus</i> rhizomes and tubers	10 days	N/A	-	76.7 ± 9.6	-	314.2 ± 16.9	-	-	-	-
Fibrianto et al. (2020)	Indonesia	<i>Camellia sinensis</i> leaves	3 days	N/A	-	23.4 ± 1.73	-	-	-	269 ± 0.71	-	-
Jakubczyk et al. (2020)	Poland	Green Tea (<i>Camellia sinensis</i>) leaves	1, 7, and 14 days	N/A	91.4 ± 0.57 ^a	-	-	-	4801.1 ± 69.2 ^a	239.1 ± 4.67 ^a	146.8 ± 3.4 ^a	-
Jakubczyk et al. (2024)	Poland	Green Tea (<i>Camellia sinensis</i>) leaves	7-14 days	N/A	91.26 ± 1.74 ^a	-	96.48 ± 4.94	-	4341.6 ± 82.63 ^a	299.6 ± 3.1 ^a	946.74 ± 222.62 ^a	-
Kaewkod et al. (2019)	Thailand	Green Tea (<i>Camellia sinensis</i>) leaves	3, 6, 9, 12, and 15 days	Caco-2 cells	-	-	-	-	-	1087.49 ± 23.12 ^a	-	-
Khaleil et al. (2020)	Saudi Arabia	Black and green tea	7 days	N/A	155	-	-	-	-	180	18	-
Li et al. (2022)	China	Green Tea (<i>Camellia sinensis</i>) leaves	7 days	N/A	-	-	37.77 ± 0.62	-	4749.39 ± 131.42	317.54 ± 5.93	104.27 ± 2.67	-
Sales et al. (2023)	Brazil	Black tea leaves	9 days	HK-2 cells	-	-	-	-	-	214 ± 2.98	-	77.03 ± 3.35
Xiong et al. (2023)	China	Bamboo leaves	14 days	N/A	-	-	-	-	1850.83 ± 138.12	303.64 ± 9.11	-	-
Xu et al. (2022)	South Korea	Green tea	6 days	N/A	95.99 ± 3.11	-	-	-	-	1360.71 ± 28.96	-	-
Zhang et al. (2023)	China	De'ang sour tea	30, 60, 90, 120 days	N/A	25.01 ± 4.8 ^b	-	-	-	-	-	-	-
Ziemlewska et al. (2021)	Poland	Yerba mate tea (<i>Ilex paraguariensis</i>) leaves	14 days	N/A	-	219.04 ± 0.11	-	174.02 ± 0.14	-	-	-	-
Ziemlewska et al. (2023)	Poland	<i>Aronia melanocarpa</i> leaves	20 days	Human fibroblasts	-	791 ± 7.4	-	128 ± 2.67	-	-	-	-
Ziemlewska et al. (2024)	Poland	Radish (<i>Raphanus sativus</i>) leaf	7, 14, and 21 days	Human fibroblasts	43.23 ± 0.12 ^a	-	96.8 ± 0.5	-	-	-	-	84.53 ± 1.16

Numerical data are presented as mean ± standard deviation. ^aData was acquired from fermentation day 6 to 9 (mostly day 7 due to the highest antioxidant power reported in previous studies), ^bData was acquired from fermentation day 30). ABTS = 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), DPPH = 2,2-diphenyl-1-picrylhydrazyl, FRAP = ferric ion reducing antioxidant power, N/A = not available ROS = reactive oxygen species, TPCs = total phenolic compounds, TFCs = total flavonoid contents.

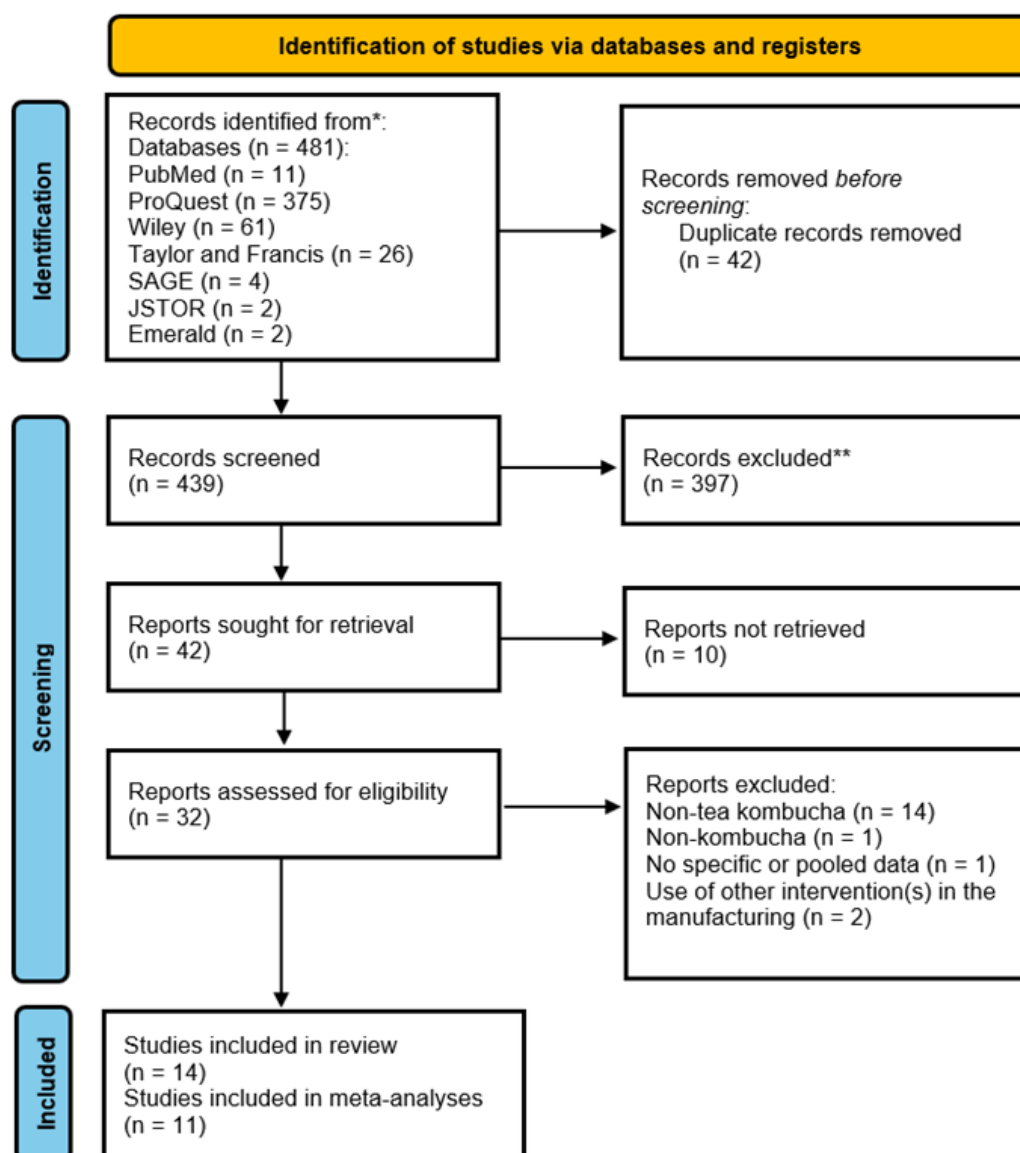


Figure 1. PRISMA flowchart

Meta-analysis

Eleven studies were included for meta-analysis, which was used to determine antioxidant capacity (radical scavenging ability), antioxidant reducing capability, oxidative stress marker, and total amount of antioxidant compounds. Regarding free radical scavenging capacity, DPPH assay showed significant difference between kombucha tea and unfermented tea, for both parameters (radical scavenging ability % and IC_{50}), as determined from three studies data. The overall mean difference of scavenging percentage was 9.84% (95% CI = 4.63 to 15.05%, $p = 0.0002$). Meanwhile,

kombucha overall IC_{50} was lower than the required concentration for unfermented tea, with overall effect of -64.54 mg/L (95% CI = -99.06 to -30.03 mg/L, $p = 0.0002$). Meanwhile, ABTS assays did not show a significant difference between tested groups, although having a similar tendency as DPPH measurements. The ABTS scavenging percentage difference was at 28.89% (95% CI = -4.58 to 62.37%, $p = 0.09$) and IC_{50} difference was at -4.82 mg/L (95% CI = -40.27 to 30.62 mg/L, $p = 0.79$). Meta-analysis data is available in Figure 2.

The current meta-analysis was expanded to determine reducing capacity

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and oxidative stress level (Figure 3), along with antioxidant compound amount (Figure 4). All analysis reflected insignificant result, with the MD or SMD as follows: FRAP (SMD = 1.38, 95% CI = -6.34 to 9.10, $p = 0.73$), intracellular ROS (MD = 7.71%, 95% CI = -1.86 to 17.29%, $p = 0.11$), TPCs (MD = -21.20 mg GAE/L, 95%

CI = -116.97 to 74.58 mg GAE/L, $p = 0.66$), and TFCs (MD = -53.54 mg GAE/L, 95% CI = -158.69 to 51.61 mg GAE/L, $p = 0.32$). Similar to radical scavenging ability assays, all measurements were having considerable heterogeneity, with I^2 values ranging from 78% to 100%.

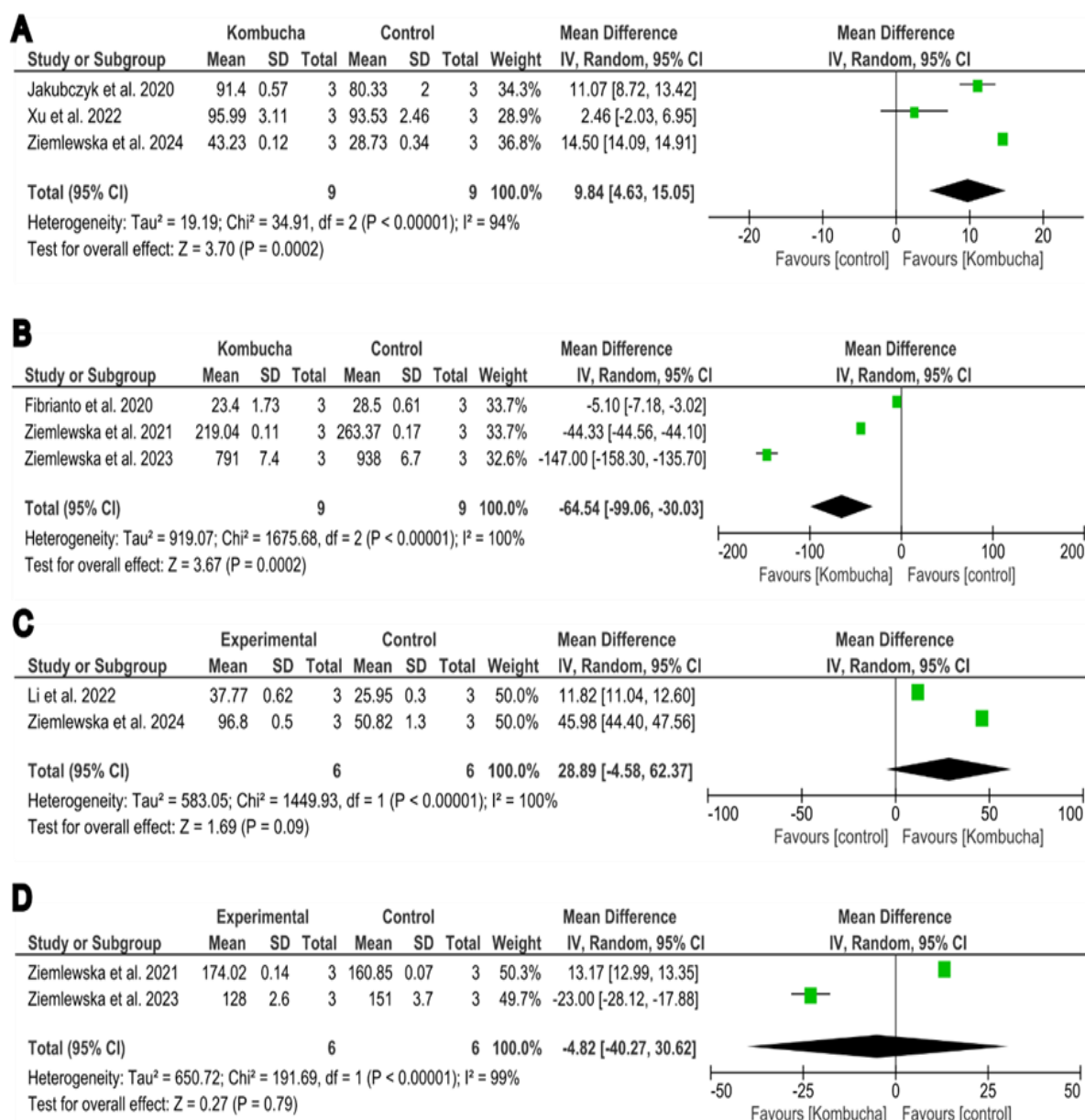


Figure 2. Statistical analysis of free radical scavenging ability. (A) DPPH scavenging percentage, (B) DPPH IC_{50} , (C) ABTS scavenging percentage, (D) ABTS IC_{50} .

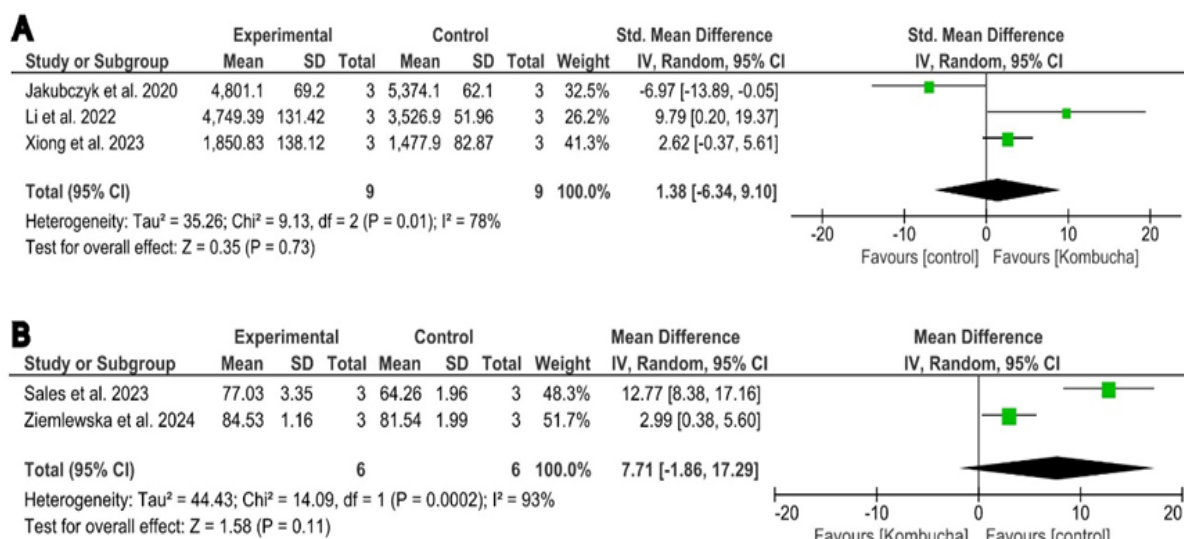


Figure 3. Antioxidant and oxidative stress parameters. (A) FRAP, (B) Intracellular ROS.

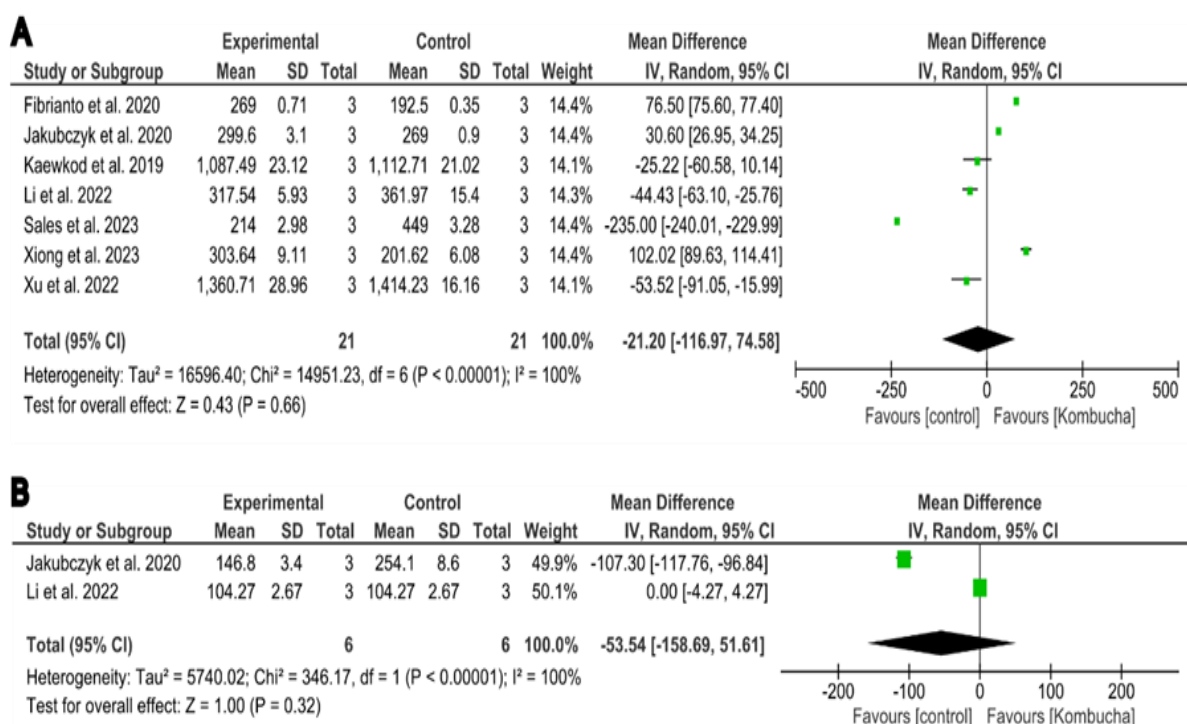


Figure 4. Total antioxidant compounds. (A) TPCs, (B) TFCs).

Risk of bias

Following quality assessment with risk of bias analysis, all studies did not report any sample size calculation (D1). However, it should be noted that all studies were conducted as triplicate, which is a very common measure in *in vitro* studies. Meanwhile, for the domains sample with similar dimensions (D2) and

standardization of procedures (D4), all studies showed a low risk of bias. In control

group (D3) assessment, three studies did not use any control, and only one study did not report a clear statistical analysis (D5). The majority of the studies showed a low risk of bias in other risk (D6) (9/14; 64.29%) (Figure 5 and Figure 6).

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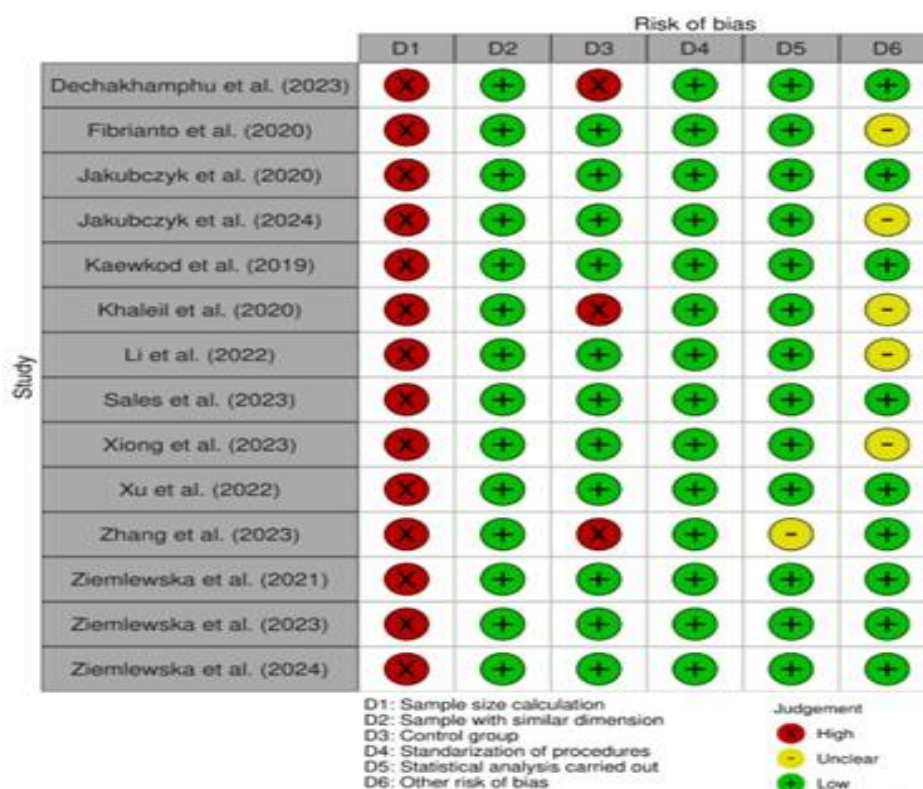


Figure 5. Traffic plot of risk of bias

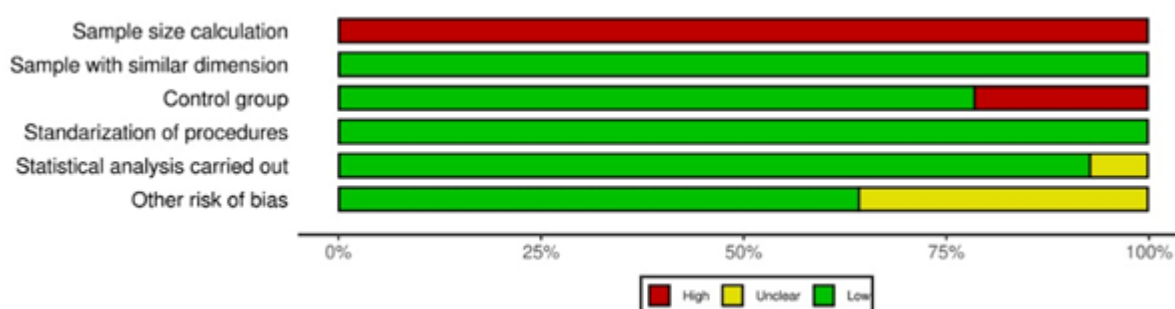


Figure 6. Summary plot of risk of bias

Discussion

Nowadays, numerous customers perceive fermentation as an organic approach to food preservation that may augment the health benefits of products, such as by improving essential amino acids, proteins, vitamins, and different metabolites (Sharma et al. 2020; García-Barón et al. 2025). From an antioxidant perspective, fermentation increases the levels of various enzymes that facilitate the production, release, and generation of bioactive chemicals from the substrate (Abbaspour 2024; Saritaş et al. 2024).

Various factors contribute to this process including elevated phytochemicals (e.g. polyphenols and flavonoids), antioxidant peptides, and antioxidant polysaccharides generated through microbial hydrolysis or biotransformation (Zhou et al. 2020; Zhao et al. 2021; Luo et al. 2024; Fitsum et al. 2025). Furthermore, fermentation can facilitate the degradation of plant cell walls, thereby aiding in the release or generation of diverse antioxidant molecules (Zhao et al. 2021, 2024).

Kombucha is an example of fermented beverages which showed growing

popularity. Despite coming from Asia, kombucha popularity is spreading to Europe and other parts of the world (Bishop et al. 2022; Andrade et al. 2025). Research on the various base types of teas, their composition, and their health-promoting qualities is limited, despite the fact that kombucha antioxidant qualities have been thoroughly examined (Gaggia et al. 2018; Jakubczyk et al. 2020; Xiong et al. 2023).

Among the bioactive compounds present in kombucha yeasts, organic acids, catechins, and polyphenols are the primary chemicals that possess antioxidant functions (Alaei et al. 2020; Abaci et al. 2022; Bortolomedi et al. 2022; das Chagas et al. 2024; Martihandini and Rochimat 2024). Although tea leaves already exhibit antioxidant properties, fermentation of tea leaves with SCOBY could further enhance the antioxidant properties via increased production of gallic acid and glucuronic acid (Rahmani et al. 2019; Chou et al. 2024). Thus, kombucha may exhibit superior antioxidant properties compared to tea (Sun et al. 2015; Jakubczyk et al. 2020). Because of resonance stabilization, phenolic compounds are able to donate hydroxyl hydrogen with ease, and this hydrogen supply gives them a great ability to scavenge DPPH (Wang et al. 2018; Muliasari et al. 2023). The other most widely used techniques are ABTS-based antioxidant capacity assays. The interaction between an antioxidant and the pre-generated ABTS^{•+} radical cation forms the basis of the ABTS/PP test (Ilyasov et al. 2018). The fading of absorption spectra characteristic peaks at 414, 417, 645, 734, and 815 nm makes it simple to quantitatively detect ABTS scavenging (Ilyasov et al. 2020).

Our review showed a trend of increment of the DPPH assay antioxidant activity, but not in ABTS assay. This may be explained by the different mechanisms involved. DPPH responds to hydrogen-donating phenolics, such as catechol and gallic acid (Brighente et al. 2007; Akar et al. 2017). Meanwhile, ABTS respond to a broader

range of antioxidants and are not limited to hydrogen-donating phenolics (Christodoulou et al. 2022). Regarding tea types, this study utilized the data from green tea when possible, since it is considered to have the highest DPPH scavenging ability (88%) compared to black tea and tea waste material following fermentation process (Jayabalan et al. 2008; Cheepchirasuk et al. 2025), along with highest overall antioxidant activities than black and red tea (Jakubczyk et al. 2020). However, this difference on antioxidant assays activity on kombucha needs further evaluation.

Our study showed a few limitations, especially in the level of certainty of evidence. The results from all of the studies were pre-preliminary (*in vitro* experiments), which showed low certainty of evidence compared to clinical trials. Despite these limitations, our study was the first to provide valuable sources of evidence about how tea-based kombucha shows potential as an antioxidant agent. Furthermore, all included studies showed significant heterogeneity in the result, which may impact study generalizability and interpretation since the included studies varied significantly in methodologies, tea types, fermentation durations, geographical variations, and analytical conditions. This significant variation presumably arises from numerous critical variables. The fermentation duration in the included research varied significantly from three to thirty days, which substantially affects the ultimate content of organic acids and phenolic compounds (Jayabalan et al. 2007). Moreover, the tea substrate is a significant variable, whether comparing teas of the same species (*Camellia sinensis*, green and black tea) (Jakubczyk et al. 2020) or distinct species (such as bamboo leaf, *Ilex paraguariensis*, and *Cyperus rotundus*) (Ziemlewska et al. 2021; Xiong et al. 2023; Dechakhamphu et al. 2023). This study also included a relatively small amount of research papers, particularly of DPPH,

which may also reduce the statistical power and robustness of the conclusions.

In conclusion, the current systematic review and meta-analysis presented a significant difference of antioxidant capacity of kombucha tea compared to control (unfermented tea) as measured by DPPH assays. However, this significance is not applicable for ABTS assays and FRAP. In addition, oxidative stress level and antioxidant compound amount also do not differ significantly. Nevertheless, due to high heterogeneity in the included studies, more studies should be conducted in more detail to determine the antioxidant activity of kombucha tea. Future research should include standardizing the reporting process, such as a full description of the SCOBY composition, the exact substrate used, and all the fermentation parameters (e.g. time, temperature, and sugar concentration). Additionally, a positive control (such as Trolox, gallic acid, and ascorbic acid) alongside the unfermented tea (negative) control should be employed to confirm assay performance. *In vivo* studies may also be conducted to ascertain the physiological significance and bioavailability of the antioxidant compounds present in kombucha.

Conflicts of interest

The authors declare no conflict of interest.

Availability of data and materials

This published article contains all of the generated or analysed data obtained during this investigation.

Authors' Contributions

Conceptualization, Z.A.; Methodology, Z.A., M.M.S., K.T., L.H., J.S., and T.P.U.; Formal analysis and results interpretation, Y.T.N.S., A.M.S., and T.P.U.; validation, Z.A., M.M.S., S.V.K., L.L., E.S., and T.P.U.; writing-original draft preparation, K.T., A.M.S., and C.T.; Writing-review and editing, M.M.S. and T.P.U.; Supervision, Z.A., L.L., L.H., J.S., and E.S. All authors

reviewed the results and approved the final version of the manuscript. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

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Supplementary

Supplementary Table 1. PRISMA 2020 Checklist

TITLE		
Title	1 Identify the report as a systematic review.	Page 1
ABSTRACT		
Abstract	2 See the PRISMA 2020 for Abstracts checklist.	Page 1
INTRODUCTION		
Rationale	3 Describe the rationale for the review in the context of existing knowledge.	Page 3-4
Objectives	4 Provide an explicit statement of the objective(s) or question(s) the review addresses.	Page 4
METHODS		
Eligibility criteria	5 Specify the inclusion and exclusion criteria for the review and how studies were grouped for the syntheses.	Page 5
Information sources	6 Specify all databases, registers, websites, organisations, reference lists and other sources searched or consulted to identify studies. Specify the date when each source was last searched or consulted.	Page 5
Search strategy	7 Present the full search strategies for all databases, registers and websites, including any filters and limits used.	Page 5
Selection process	8 Specify the methods used to decide whether a study met the inclusion criteria of the review, including how many reviewers screened each record and each report retrieved, whether they worked independently, and if applicable, details of automation tools used in the process.	Page 5
Data collection process	9 Specify the methods used to collect data from reports, including how many reviewers collected data from each report, whether they worked independently, any processes for obtaining or confirming data from study investigators, and if applicable, details of automation tools used in the process.	Page 5
Data items	10a List and define all outcomes for which data were sought. Specify whether all results that were compatible with each outcome domain in each study were sought (e.g. for all measures, time points, analyses), and if not, the methods used to decide which results to collect.	Page 6
	10b List and define all other variables for which data were sought (e.g. participant and intervention characteristics, funding sources). Describe any assumptions made about any missing or unclear information.	Page 6
Study risk of bias assessment	11 Specify the methods used to assess risk of bias in the included studies, including details of the tool(s) used, how many reviewers assessed each study and whether they worked independently, and if applicable, details of automation tools used in the process.	Page 6
Effect measures	12 Specify for each outcome the effect measure(s) (e.g. risk ratio, mean difference) used in the synthesis or presentation of results.	Page 7
Synthesis methods	13a Describe the processes used to decide which studies were eligible for each synthesis (e.g. tabulating the study intervention characteristics and comparing against the planned groups for each synthesis (item #5)).	Page 7
	13b Describe any methods required to prepare the data for presentation or synthesis, such as handling of missing summary statistics, or data conversions.	Page 7
	13c Describe any methods used to tabulate or visually display results of individual studies and syntheses.	Page 7
	13d Describe any methods used to synthesize results and provide a rationale for the choice(s). If meta-analysis was performed, describe the model(s), method(s) to identify the presence and extent of statistical heterogeneity, and software package(s) used.	Page 6-7
	13e Describe any methods used to explore possible causes of heterogeneity among study results (e.g. subgroup analysis, meta-regression).	N/A
	13f Describe any sensitivity analyses conducted to assess robustness of the synthesized results.	N/A

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Reporting bias assessment	14	Describe any methods used to assess risk of bias due to missing results in a synthesis (arising from reporting biases).	N/A
Certainty assessment	15	Describe any methods used to assess certainty (or confidence) in the body of evidence for an outcome.	N/A
RESULTS			
Study selection	16a	Describe the results of the search and selection process, from the number of records identified in the search to the number of studies included in the review, ideally using a flow diagram.	Page 8
	16b	Cite studies that might appear to meet the inclusion criteria, but which were excluded, and explain why they were excluded.	Figure 1
Study characteristics	17	Cite each included study and present its characteristics.	Page 8
Risk of bias in studies	18	Present assessments of risk of bias for each included study.	Page 9-10, Figure 5
Results of individual studies	19	For all outcomes, present, for each study: (a) summary statistics for each group (where appropriate) and (b) an effect estimate and its precision (e.g. confidence/credible interval), ideally using structured tables or plots.	Page 8, Table 1
Results of syntheses	20a	For each synthesis, briefly summarise the characteristics and risk of bias among contributing studies.	Page 9-10
	20b	Present results of all statistical syntheses conducted. If meta-analysis was done, present for each the summary estimate and its precision (e.g. confidence/credible interval) and measures of statistical heterogeneity. If comparing groups, describe the direction of the effect.	Page 8
	20c	Present results of all investigations of possible causes of heterogeneity among study results.	N/A
	20d	Present results of all sensitivity analyses conducted to assess the robustness of the synthesized results.	N/A
Reporting biases	21	Present assessments of risk of bias due to missing results (arising from reporting biases) for each synthesis assessed.	N/A
Certainty of evidence	22	Present assessments of certainty (or confidence) in the body of evidence for each outcome assessed.	N/A
DISCUSSION			
Discussion	23a	Provide a general interpretation of the results in the context of other evidence.	Page 11-12
	23b	Discuss any limitations of the evidence included in the review.	Page 12-13
	23c	Discuss any limitations of the review processes used.	Page 12-13
	23d	Discuss implications of the results for practice, policy, and future research.	Page 13
OTHER INFORMATION			
Registration and protocol	24a	Provide registration information for the review, including register name and registration number, or state that the review was not registered.	Page 5
	24b	Indicate where the review protocol can be accessed, or state that a protocol was not prepared.	N/A
	24c	Describe and explain any amendments to information provided at registration or in the protocol.	N/A
Support	25	Describe sources of financial or non-financial support for the review, and the role of the funders or sponsors in the review.	Page 13
Competing interests	26	Declare any competing interests of review authors.	Page 13
Availability of data, code and other materials	27	Report which of the following are publicly available and where they can be found: template data collection forms; data extracted from included studies; data used for all analyses; analytic code; any other materials used in the review.	Page 14

From: Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ* 2021;372:n71. doi: 10.1136/bmj.n71. This work is licensed under CC BY 4.0. To view a copy of this license, visit <https://creativecommons.org/licenses/by/4.0/>